



US006518414B1

(12) **United States Patent**
MacLennan

(10) **Patent No.:** **US 6,518,414 B1**
(45) **Date of Patent:** **Feb. 11, 2003**

(54) **MOLECULAR CLONING AND EXPRESSION OF G-PROTEIN COUPLED RECEPTORS**

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(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days.

(21) Appl. No.: **09/225,024**

(22) Filed: **Jan. 4, 1999**

Related U.S. Application Data

(63) Continuation-in-part of application No. 08/760,936, filed on Dec. 6, 1996, now Pat. No. 5,856,443, which is a continuation of application No. 08/196,989, filed on Feb. 15, 1994, now Pat. No. 5,585,476.

(51) **Int. Cl.**⁷ **C12N 15/12**

(52) **U.S. Cl.** **536/23.5; 435/69.1**

(58) **Field of Search** 435/7.1, 7.2; 475/69.1; 530/350; 536/23.5

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Supplemental Information Disclosure Statement Under 37 CFR §§1.97 and 1.98 dated Mar. 14, 2002.

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(57) **ABSTRACT**

The subject invention pertains to cloning and expression of novel cDNAs which encode members of the G-protein coupled receptor superfamily of proteins. Polynucleotides which encode mammalian H218 protein are described. The invention also concerns methods for screening for ligands of H218 protein. The proteins and peptides of the subject invention can also be used to produce antibodies which can bind to the subject proteins. The polynucleotide molecules, proteins, and antibodies of the subject invention can be used in both diagnostic and therapeutic applications.

2 Claims, 13 Drawing Sheets

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-155 CCCCCCCCCCTCGAGCACAGCCAACAGTCACCAAAAGTCAGCCACTGGCTGTCCCGG
-95 GCGCAGACGCCAAGCCACTCAGGCCAGGCGACCCCTGGCCGCGCTAGCCAGTGCT
-35 CAGTCCCATGGCCCCGCGGCCACTGAGCCGAGCGTGGCGGTTTATACTCAGAGTAC
      MetGlyGlyLeuTyrSerGluTyr      8
25 CTCAATCCTGAGAAGGTTCAGGAACACTACAAATTACACCAAGGAGACGCTGGACATGCAG
   LeuAsnProGluLysValGlnGluHisTyrAsnTyrThrLysGluThrLeuAspMetGln 28
85 GAGACGCCCTCCCGCAAGGTGGCCTCCGCCCTTCATCATCATTTTATGCTGTGCCATCGTG
   GluThrProSerArgLysValAlaSerAlaPheIleIleLeuCysCysAlaIleVal 48
145 GTGGAGAACCTTCTGCTGCTAATCGCAGTGGCCAGGAACAGCAAGTTCCACTCAGCCATG
   ValGluAsnLeuLeuValLeuIleAlaValAlaArgAsnSerLysPheHisSerAlaMet 68
205 TACCTGTTCTCGGCAACCTGGCAGCCTCCGACCTGCTGGCAGGGCGTGGCTTCGTGGCC
   TyrLeuPheLeuGlyAsnLeuAlaAlaSerAspLeuLeuAlaGlyValAlaPheValAla 88
265 AACACCTTGCTCTCCGGACCTGTACCCCTGTCTTAACCTCCCTTGAGTGGTTTGCCCGA
   AsnThrLeuLeuSerGlyProValThrLeuSerLeuThrProLeuGlnTrpPheAlaArg 108
325 GAGGGTTCAGCCTTCATCACGCTCTCTGCCCTCGGCTCTCAGCCTCCTGGCCATGGCCATC
   GluGlySerAlaPheIleThrLeuSerAlaSerValPheSerLeuLeuAlaIleAlaIle 128
385 GAGAGACAAGTGGCCATCGCCAAGGTCAAGCTCTACGGCAGTGACAAAAGCTGTGGAATG
   GluArgGlnValAlaIleAlaLysValLysLeuTyrGlySerAspLysSerCysArgMet 148
445 TTGATGCTCATTTGGGGCCTCTTGGCTGATATCGCTGATTCTGGGTGGCTTGGCCATCCTG
   LeuMetLeuIleGlyAlaSerTrpLeuIleSerLeuIleLeuGlyGlyLeuProIleLeu 168
505 GGCTGGAATTGCTGGACCATCTGGAGGCTTGCTCCACTGTGCTGCCCTCTATGCTAAG
   GlyTrpAsnCysLeuAspHisLeuGluAlaCysSerThrValLeuProLeuTyrAlaLys 188
565 CACTATGTGCTCTGCGTGGTCACCATCTTCTCTGTCACTTACTGGCTATCGTGGCCTTG
   HisTyrValLeuLeuCysValValThrIlePheSerValIleLeuLeuAlaIleValAlaLeu 208

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FIG. 1A

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625	TACGTCCGAATCTACTTCGTAGTCCGCTCAAGCCATGCGGACGTTGCTGCTCCTCAGACG TyrValArgIleTyrPheValValArgSerSerHisAlaAspValAlaGlyProGlnThr	228
685	CTGGCCCTGCTCAAGACAGTCACCATCGTACTGGGTGTTTTCATCATCTGCTGGCTGCCG LeuAlaLeuLeuLysThrValThrIleValLeuGlyValPheIleIleCysTrpLeuPro	248
745	GCTTTTAGCATCCTTCTCTTAGACTCTACCTGTCCCGTCCGGCCTGTCCCTGTCCCTCTAC AlaPheSerIleLeuLeuLeuAspSerThrCysProValArgAlaCysProValLeuTyr	268
805	AAAGCCCATATTCTTTCCTTCGCCACCCCTCAACTCTCTGCTCAACCCCTGTCTCTAT LysAlaHisTyrPhePheAlaPheAlaThrLeuAsnSerLeuLeuAsnProValIleTyr	288
865	ACATGGCGTAGCCGGACCTTCGGAGGGAGGTACTGAGGCCCTGCTGTGCTGGCGGCAG ThrTrpArgSerArgAspLeuArgArgGluValLeuArgProLeuLeuCysTrpArgGln	308
925	GGGAAGGAGCAACAGGCGCAGAGGTGGGAACCCCTGGTCACCGACTCCTGCCCTCCGC GlyLysGlyAlaThrGlyArgArgGlyGlyAsnProGlyHisArgLeuLeuProLeuArg	328
985	AGCTCCAGCTCCCTGGAGAGAGGCTTGCCATATGCCCTACATCGCCCAACATTCTGGAGGGC SerSerSerLeuGluArgGlyLeuHisMetProThrSerProThrPheLeuGluGly	348
1045	AACACAGTGGTCTGAGGGGAAATGTGAAGTCTGTAAACCAAGCCACAGAGAGAGCTCT AspThrValVal	352

FIG. 1B

1105 GTGGGAGAGACCAGGTGACCTCATCATGTCTCCCTCAGTGCCACAGGCTCTGGAGGAACTGA
 1165 CCACGGCTCATAGGTCAGGTGGCCAAACGGAGGCACGTGACTAATCAGATTGTAGTACTGTG
 1225 ACTGTGGGACCAATTAAGGGCTAGGGGACACAGCGCTCGAGTTAGGGCTAGACATTT
 1285 GCCACTTGGTACATAGGGTGTCCGCACTCTGTCTCTTCTATCTTCCAGCTTCCCGGTTCC
 1345 CTTCCTGCCCTCCTCTTTAAGGGCTCTCTACATAGCCCCGGCTGGCTAGAGCTTGCTG
 1405 TGCAGACCAAGCTGACCTGGACCTCCAGAGATAGATCAACTAACTGTGTCTGAGTGCT
 1465 GGGATTTTAAAGCCGTGTGCCCCACACCCGGCTCTGCCACCTTCCAGAGCAATCTTA
 1525 GGCCACTTGTGAGGAAACACTCTCCCCAGAGGACCCCAAGCCCTCTTCCCCTCTCTCTG
 1585 AGGCCCTGAATCCACAGCTTCCCCATTATCAACTGCTGCTTCTTCCCCTTCTCTCTG
 1545 TTCAGGGGAAACCACTGTGGGGGAGGGGCTCTGGATCCAGTTTATGCTCAG
 1605 ATCTCACTGAGCACCTTGCTTATTTGGGAGCAGAGAGGAATCAGCTGAGGCAGTGTGGG
 1665 CAGATGTTGAGGAGAAATTTGGGCTTCCCTGGTGAGAAACTCTAGGGGAGGCCGTTGGTTAT
 1725 TCCTGGAACCCAGCCTCTCTCCCCACGAACTCTTCACACCCGAGCCCTTGAGCTGGATGC
 1785 AAAGGCTGCTTTCAATTTGTCCTTTGTAGTTTGTGTTTGTGTTTGTGTTTAAATT
 1845 GGGACAGGATCTCAGGTACCCAGGCTGGCCTCCGACTCAGTATGTAGCCAAGGCTGGCT
 1905 TTGGACTTCTGACCCCTCCCTGCCCTCCGCTTCTGGAGTGCAGGTATTACAAGGTGTACCAC
 1965 CACCACCACCAACCAACAACAACAACACACCTGTCTTGAAAACTATCATGA
 2025 ATGACATGGTTCACATAGCCTTGGGTGGCCAGGACATCCCGGATACTCTTATGGCATCT
 2085 TCCTTGAAGGACTTTGCTAAATCCTGTGGAGAGTAGAAATCCAATACGGTACAAACGG
 2145 TATTTATGTGTGTCTGTATCAGTGTGGGCTGTGTGACCTCCTATCCCAGTGTGGGTGC
 2205 TGTCTGACCTCTTATGTGCACATCCGTGTCAAGACTGTCTAGAGAGATGACGGGGTGTG
 2265 TGTGCTTGTGGGGTCTAGCCATGATCAGGCCCTCCTGGGAATTGCTGAATCATCTCTCCC
 2325 ACACACAGACACACACCTCCGCCCTTAAAGAAATGTGTGAAAGAAAGGCTGAGGAAGGG
 2385 AGATTTGGGAGGCAAGGAGCCAGTCGGGAGTGTGCTCCCCCTCATACAGCTTCCCCAGATG
 2445 TCCCCCTTGTGCTGGAAACCCAGAACTGGGCCCAATAAACAGTTCAATTCTCTTGAAAA
 2505 AAA

FIG. 1C

FIG. 2A

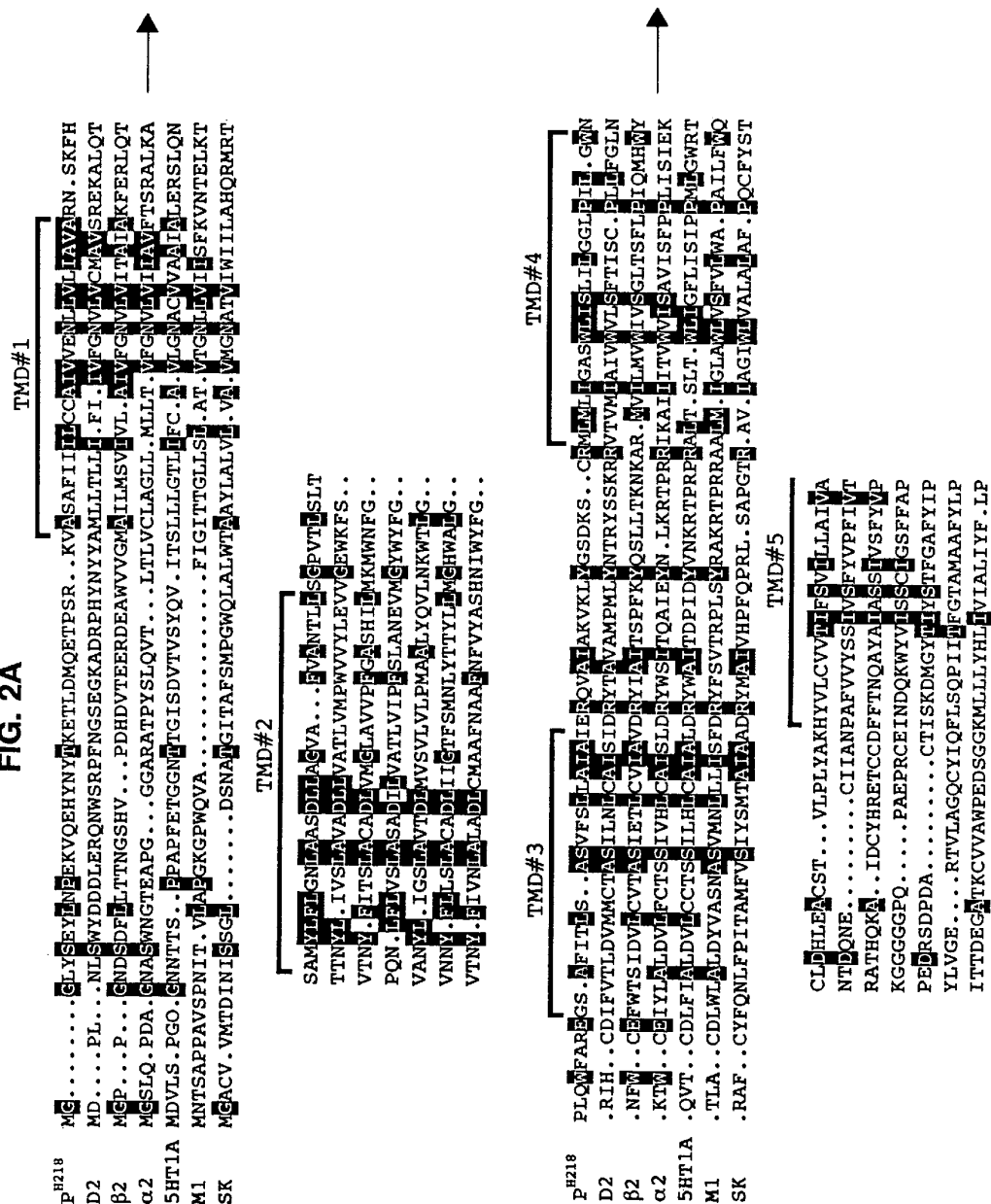
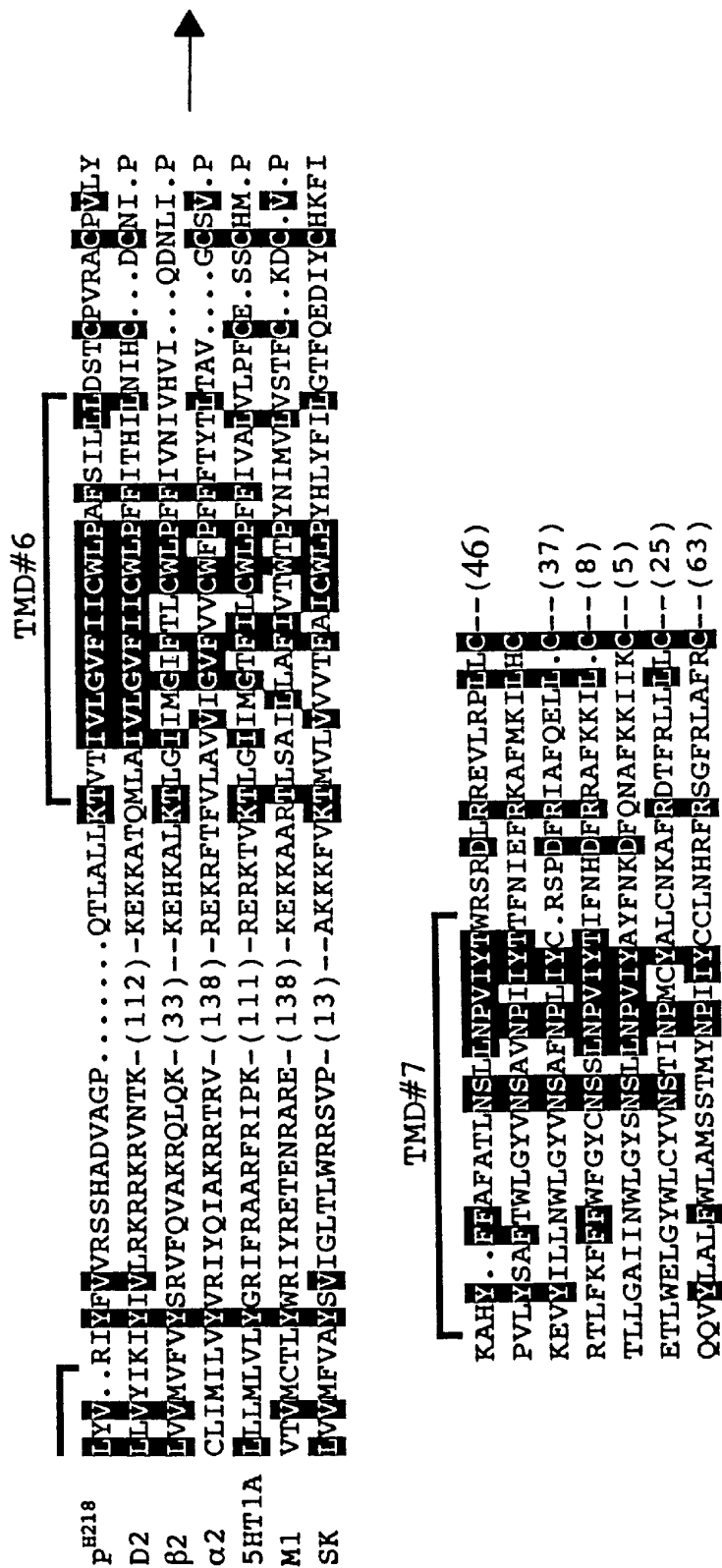


FIG. 2B

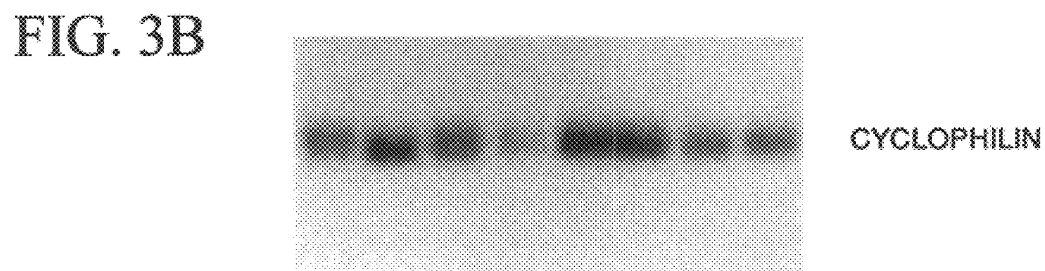
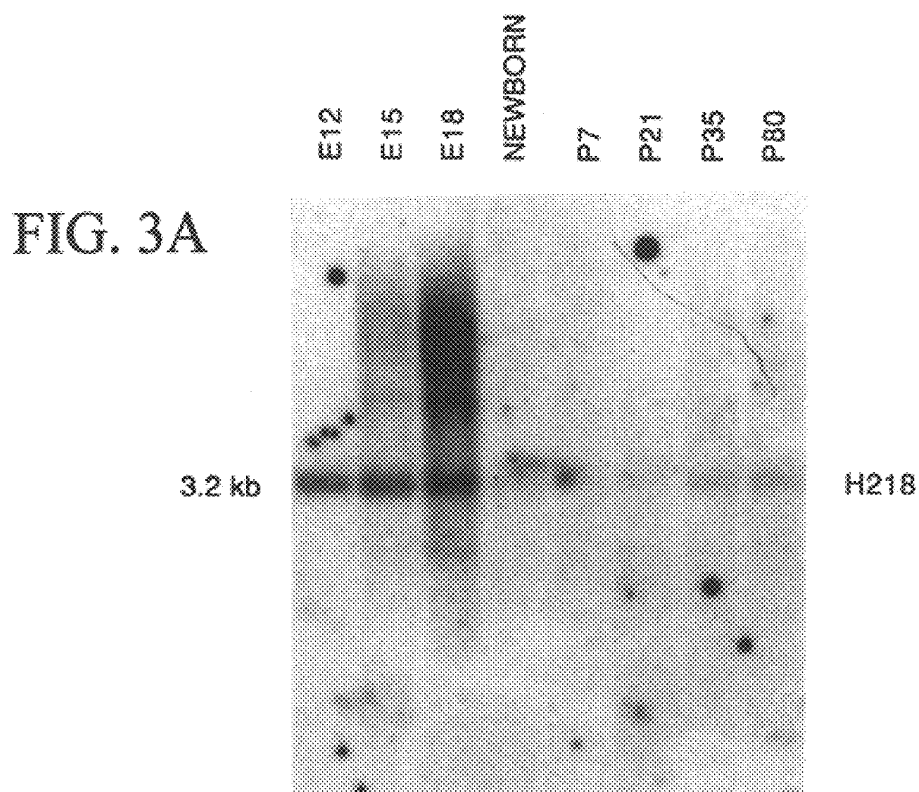


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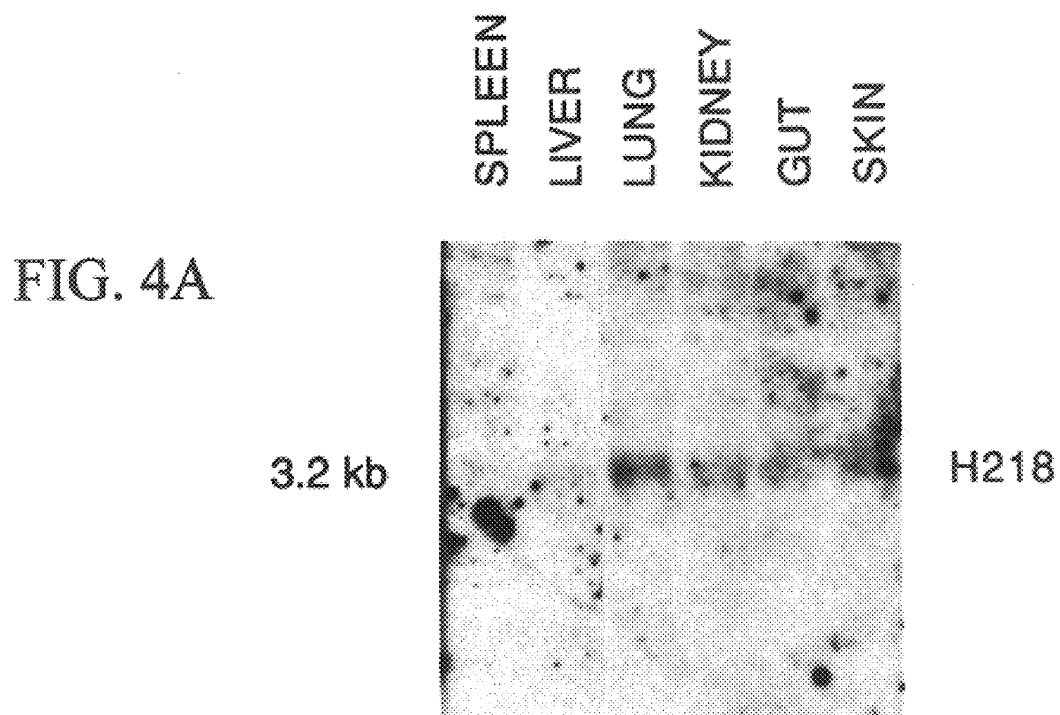
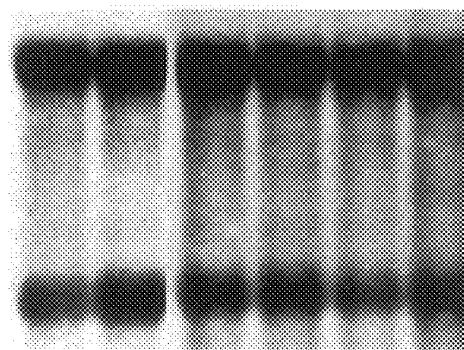


FIG. 4B



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FIG. 5A

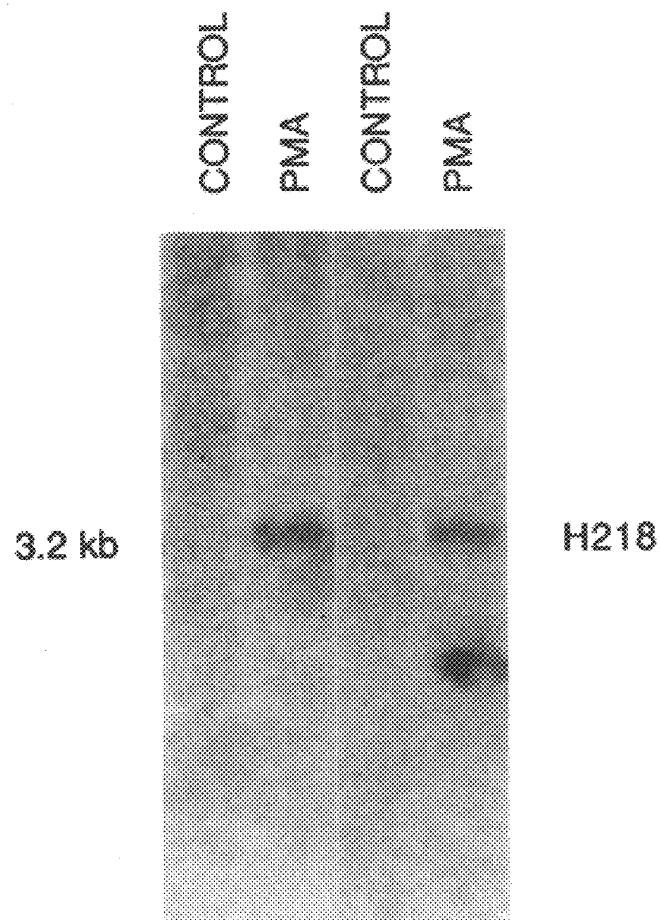
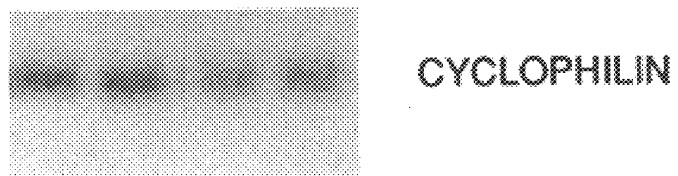


FIG. 5B



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FIG. 6A

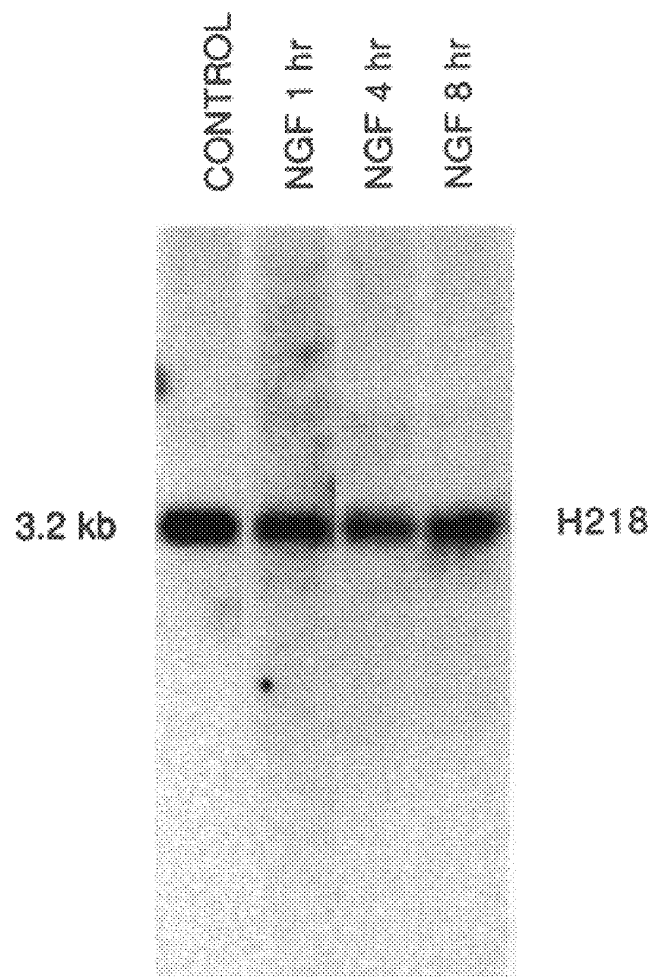
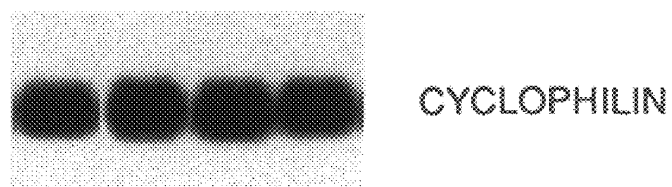


FIG. 6B



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-260      TTTGCTGGTCTCCGTCACTCGCCGACAGCAGCAAGATCGGGATCGCGGGTGTAG
-206  ACCCGGAGCCCGGGACGAGCTTCGTCCTCCGCTTGAGCGAGGCTGCTGTTCTCGGAGG
-146  CCTCTCCAGCCCAAGGAAATACTACATAAAAGCATCGGATGCTTGCTGACCTGGCCTT
-86   GCTGTAACTGAAGGCTCGCTCAACCTCGCCCTTAGCGTTTGTCTGGAGAAGTACCACCC
-26   CGGGCTCCTGGGACACAGTTGCGGCTATGGTGTCCTCCACCATCCAGTGGTTAAG
      MetValSerSerThrSerIleProValValLys  11

34   GCTCTCCGCAGCCAAAGTCTCCGACTATGGCAACTATGATATCATAGTCCGGCATTACAAC
      AlaLeuArgSerGlnValSerAspTyrGlyAsnTyrAspIleIleValArgHisTyrAsn  31

94   TACACAGGCAAGCTGAACATCGGAGTGGAGAAGGACCATGGCATTAACACTGACTTCAGTG
      TyrThrGlyLysLeuAsnIleGlyValGluLysAspHisGlyIleLysLeuThrSerVal  51

154  GTGTTCAATCTCATCTGCTGCTTGATCATCCTAGAGAAATATATTGCTGTGCTAACTATT
      ValPheIleLeuIleCysCysLeuIleIleLeuGluAsnIlePheValLeuLeuThrIle  71

214  TGGAAACCAAGAAGTTCACCGGCCCATGTAATACTATTTATAGCAACCTAGCCCTCTCG
      TrpLysThrLysLysPheHisArgProMetTyrTyrPheIleGlyAsnLeuAlaLeuSer  91

274  GACCTGTTAGCAGGAGTGGCTTACACAGCTAACCTGCTGTGTGCTGGGGCCACCACTAC
      AspLeuLeuAlaGlyValAlaTyrThrAlaAsnLeuLeuLeuSerGlyAlaThrThrTyr  111

334  AAGCTCACACCTGCCAGTGGTTTCTGCGGGAAGGAAGTATGTTGTGGCTCTGTCTGCC
      LysLeuThrProAlaGlnTrpPheLeuArgGluGlySerMetPheValAlaLeuSerAla  131

394  TCAGTCTTCAGCCTCCTTGCTATCGCCATTGAGCGCTACATCACCATGCTGAAGATGAAA
      SerValPheSerLeuLeuAlaIleAlaIleGluArgTyrIleThrMetLeuLysMetLys  151

454  CTACACAACGGCAGCAACAGCTCGCGCTCCTTTCTGCTGATCAGTGCCTGCTGGGTCATC
      LeuHisAsnGlySerAsnSerSerArgSerPheLeuLeuIleSerAlaCysTrpValIle  171

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FIG. 7A

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514 TCCCTCATCTGGGTGGGTGCCCATCATGGGCTGGAACATGCATCAGCTCGCTGTCCAGC
 SerLeuIleLeuGlyGlyLeuProIleMetGlyTyrAsnCysIleSerSerLeuSerSer 191
 594 TGCTCCACCGTGTCCCGCTCTACCACAAGCACTATATCTCTTCTGTGCACACCGTCTTC
 CysSerThrValLeuProLeuTyrHisLysHisTyrIleLeuPheCysThrThrValPhe 211
 654 ACCCTGCTCCTGCTTCCATCGTCATCTCTACTGCAGGATCTACTCCTTGGTGAGGACT
 ThrLeuLeuLeuSerIleValIleLeuTyrCysArgIleTyrSerLeuValArgThr 231
 714 CGAAGCCCGCCCTGACCTTCCGCAAGAACAATCTCCAAGGCCAGCCGCGAGTCCGAGAAG
 ArgSerArgArgLeuThrPheArgLysAsnIleSerLysAlaSerArgSerSerGluLys 251
 774 TCTCTGGCCTTGCTGAAGACAGTGTATCTCTCTGAGTGTCTTCTTCAATGCTGCTGGGCC
 SerLeuAlaLeuLeuLysThrValIleIleValLeuSerValPheIleAlaCysTrpAla 271
 834 CCTCTCTTCATCTACTACTTTTAGATGTGGGTGCAAGGGAAGACCTGTGACATCCTG
 ProLeuPheIleLeuLeuLeuLeuAspValGlyCysLysAlaLysThrCysAspIleLeu 291
 894 TACAAAGCAGAGTACTTCTGTTCTGGTGTGTGTAACCTCAGGTACCAACCCCATCATC
 TyrLysAlaGluTyrPheLeuValLeuAlaValLeuAsnSerGlyThrAsnProIleIle 311
 954 TACACTCTGACCAATAAGGAGATGCGCGCGGCTTCATCAGGATCATATCTTGTGCAAA
 TyrThrLeuThrAsnLysGluMetArgArgAlaPheIleArgIleIleSerCysCysLys 331
 1114 TGCCCCAACGGAGACTCCGCTGGCAAAATTCAGAGGCCCATCATCCCGGCGCATGGAATTT
 CysProAsnGlyAspSerAlaGlyLysPheLysArgProIleIleProGlyMetGluPhe 351
 1194 AGCCGCAGCAAAATCAGACAACTCCTCCACCCCAAGAGGATGATGGGACAAATCCAGAG
 SerArgSerLysSerAspAsnSerSerHisProGlnLysAspAspGlyAspAsnProGlu 371
 1254 ACCATTATGTCTTCTGGAAACGTCAATCTTCTTAAACCGGAAGCTGTTGATACTG
 ThrIleMetSerSerGlyAsnValAsnSerSerSer*** 383

FIG. 7B

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1314 TTGATTCTGGCTTCATCACTCACTACCCCTAGCATTTCAAAAACATCTCTTTCTCCACT
1374 GCTGCAAGGAAGACAGCCGGAGCCCTGAGAGAGGGAGGAGAAATGTGCGGCTT
1434 GGTGATACCATGTGTAGGTATGATTATGAACAATGCCCTGGGAAGGGTGGAGAT
1494 CAGATCTGCCCTGCAGAGGGTTTCCTGCCCCCTCCTAATCTCTTCACTTCTTCAAGTCGTT
1554 TCTGTTTATCCCCCATACTCTTTTCTTCTTCTCCTGTTTCTCTCATTTCCCCCTTCTCTACC
1614 ATCGCTTCTTTCTCTTCTTTTAAATTTTGGGGCAACAAGGAATCCCAACAATGGA
1674 TATTGTGGAAACATAGTGTGCTGAATGACGGCAAGAAATGGTGGTAATAATCAAAAGATAAAT
1734 TAACTTCATAAGACTGCTATTCTGAAATGCCAAACAATCTTGTAAGTCAGGACTGATAAAA
1794 TGGAGCAATCAGACATTTTCAGATGCCCGTCAATGTAAATCACCTACTTGAACATTTGTAT
1854 GCAATACATTCACACACAAAAGCAAAATACGTAGCCTTATTTGAACAATACCTGAACATCAT
1914 AAATACTCATGGTTTCACTCTGTCCAGGCGCCTAAGGACTATGCTGCTGTAAATACAGGAA
1974 AACACAGCGGATGCCCTCTATTAAATGTCACTCAAGAAAGTCTCTTGTAAACGTAAA
2034 GGCAAAACACATGTAGCTACTGAGCTATGACTGTCCTTGGTCACACTCTATGGGAAAAACA
2094 CCGGACTCCAC

FIG. 7C

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5'-ccgcagacgctagccctgctcaagacgggcaccatcgtgctaggcgtc

P Q T L A L L K T V T I V L G V

tttatcgtctgctggctgcccgccttcagcatcctccttctggactatgcctgtcccgtc

F I V C W L P A F S I L L D Y A C P V

cactcctgcccgatcctctacaaagcccactacttttgcgctctccaccctg -3'

H S C P I L Y K A H Y F F A V S T L

FIGURE 8

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MOLECULAR CLONING AND EXPRESSION OF G-PROTEIN COUPLED RECEPTORS

CROSS-REFERENCE TO A RELATED APPLICATION

This application is a continuation-in-part of application Ser. No. 08/760,936, filed on Dec. 6, 1996, now U.S. Pat. No. 5,856,443, which is a continuation of application Ser. No. 08/196,989, filed on Feb. 15, 1994, now U.S. Pat. No. 5,585,476.

This invention was made with government support under the National Institute on Drug Abuse grant number DA07244. The government has certain rights in this invention.

BACKGROUND OF THE INVENTION

The development of multicellular organisms requires the orchestration of many precisely coordinated events involving cell-type specific growth, proliferation, differentiation, migration, and cell death. Not surprisingly, intercellular communication plays critical roles in these processes. Although the molecular mechanisms involved in this communication are in general poorly understood, this research field is characterized by increasingly rapid progress initiated by the realization that viral oncogenes are, in many cases, transformed versions of cellular genes (proto-oncogenes) that participate in the intercellular communication directing development. Furthermore, it has been established that many non-viral forms of cancer also result from transformation of genes involved in signal transduction (e.g. growth factors, growth factor receptors, and transcription factors).

A large number of mammalian growth factor receptors have been cloned and many are recognized proto-oncogenes (Yarden and Ullrich, 1988). Most of these cloned receptors are members of a superfamily of integral membrane proteins with intrinsic, growth factor-inducible, tyrosine kinase activity. An extensive research literature now documents the critical roles these receptors play in cell proliferation, differentiation, and malignant transformation. However, multiple lines of evidence suggest that members of the G-protein coupled receptor (GPR) superfamily may also participate in mammalian development and oncogenesis. For example, both the yeast *S. cerevisiae* and the slime mold *D. discoideum* express GPRs that regulate cell differentiation (Devreotes, 1989; Sprague, 1991). In addition, mammalian mitogenesis and cell proliferation are affected by several peptides and neurotransmitters which are known to interact with GPRs (Hanley, 1989; Zachary et al., 1987).

Perhaps the most direct evidence linking GPRs with ontogeny and cancer has been provided by the ectopic expression of GPRs in tissue culture cells. Thus, the *mas* oncogene encodes a putative GPR (p^{mas}) and leads to malignant transformation when transfected into NIH3T3 mouse fibroblasts cells (Young et al., 1986). In addition, several serotonin and muscarinic acetylcholine receptors (all GPRs) also produce this malignant transformation if ectopically expressed in NIH3T3 cells and stimulated by their respective ligands (Gutkind et al., 1991; Julius et al., 1989; Julius et al., 1990). While these data illustrate that GPRs can greatly influence cell proliferation and morphology, the GPRs that were studied are unlikely to be involved in these processes in vivo because they reside in fully differentiated, postmitotic cells such as neurons where serotonergic receptors, muscarinic receptors, and most likely p^{mas} regulate the changing electrical properties of neuronal membranes involved in neurotransmission. However, these data

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support the possibility that other GPRs are expressed in vivo in immature cells where they regulate proliferation and differentiation. Furthermore, these data suggest that some forms of cancer may result from mutations or viral infections that lead to improper functioning, activation, or expression of such GPRS. Thus, identification and characterization of such receptors should significantly advance both the study of normal development as well as the search for diagnostic and therapeutic tools in oncology.

BRIEF SUMMARY OF THE INVENTION

The subject invention concerns the cloning and sequencing of cDNAs and the proteins encoded by those cDNAs. The cDNAs encode novel polypeptides that are members of the G-protein coupled receptor (GPR) superfamily. The proteins encoded by the DNAs of the subject invention are involved in the regulation of cell proliferation and/or differentiation in vivo. The subject protein receptors are endogenously expressed in various tissues and cell lines.

Specifically, the subject invention concerns the cloning and sequencing of a rat cDNA (H218) that encodes a novel GPR designated p^{H218} . Further included in the subject invention are mammalian homologs, including the human homolog of the H218 cDNA. The H218 cDNA was used to determine that H218 mRNA is expressed in all developing organs tested and in seven out of seven cell lines tested. In addition, in the brain, H218 mRNA is much more highly expressed during a period of extensive proliferation and differentiation (embryogenesis) than a period of very limited cell proliferation and differentiation (adulthood), suggesting that p^{H218} does not function as a neurotransmitter receptor. Rather, p^{H218} functions as a growth factor ligand receptor.

The subject invention further concerns antibodies from animals immunized with peptides derived from p^{H218} GPR. Purified antibody made against one of the peptides recognizes a protein having an apparent molecular weight of 50–55 kDa as determined by Western blot analysis.

The subject invention also concerns cDNA of the rat-*edg* gene. Rat-*edg* cDNA encodes a GPR, $p^{rat-edg}$. The $p^{rat-edg}$ can be activated by some of the same ligand(s) that activate p^{H218} . By identifying compounds that specifically activate or inhibit this class of receptors one can develop unique, pharmaceutical therapies that effectively treat some forms of cancer.

A further aspect of the subject invention concerns polynucleotide molecules that are antisense to mRNA of H218 and rat-*edg*. The antisense polynucleotide molecules can be used to reduce or inhibit the expression of the subject protein by binding to the complementary mRNA transcripts.

The subject invention also concerns methods of use for the polynucleotide sequences, the encoded proteins, peptide fragments thereof, polynucleotide molecules that are antisense to the H218 and rat-*edg* sequences, and antibodies that bind to the proteins and peptides. Such use includes diagnostic and therapeutic applications of the subject invention.

BRIEF DESCRIPTION OF THE DRAWINGS

FIGS. 1A–1C shows the nucleotide (SEQ ID NO.1) and deduced amino acid sequence (SEQ ID NO.2) of H218 cDNA. The sequence was compiled from that of “H2” cDNA (nucleotides 16 to 2505) and “18” cDNA (nucleotides-155 to 288) which are identical throughout the region of overlap. A black box highlights the optimal consensus sequence for translation initiation. A potential polyadenylation signal is double-underlined and a consensus

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sequence associated with mRNA instability is boxed. Repetitive nucleic acid sequences in the 3Q untranslated region are underlined. An arrow designates a predicted N-glycosylation site. A consensus sequence for proline directed kinases is underlined with a broken line. Brackets below the amino acid sequence indicate possible nucleotide binding site components in the carboxy-terminal and "third cytoplasmic loop" domains respectively.

FIGS. 2A and 2B shows a comparison of p^{H218} (SEQ ID NO.2) with other G-protein coupled receptors. Black boxes highlight residues identical to p^{H218} residues. D2=D2 dopaminergic receptor (SEQ ID NO.9); β2=β2 adrenergic receptor (SEQ ID NO.10); α2=α2 adrenergic receptor (SEQ ID NO.11); 5HT1A=1A serotonergic receptor (SEQ ID NO.12.); M1=M1 muscarinic receptor (SEQ ID NO.13); SK=substance K receptor (SEQ ID NO.14). The numbers in parentheses indicate the number of omitted residues.

FIGS. 3A and 3B shows an X-ray autoradiograph of a Northern blot illustrating the ontogenic regulation of H218 mRNA levels in the rat brain: Poly-A RNA was extracted from whole rat brain at embryonic days 12, 15, 18, Birth, postnatal days 7, 21, 35, and 80 (adult). The resulting blot was probed for H218 mRNA (panel A), stripped, and then probed with a cyclophilin cDNA (panel B) to control for variation in extraction, loading, and transfer (brain cyclophilin mRNA levels are reported to be stable from E12 to adult). The relative intensity of the cyclophilin bands have consistently paralleled results obtained from probing the same blots with an oligo-dT probe designed to hybridize to all mRNA poly-A tails.

FIGS. 4A and 4B shows an X-ray autoradiograph of a Northern blot illustrating the distribution of H218 mRNA in various tissues of the postnatal day 14 rat. Approximately 20 μg of total RNA was loaded per lane. The blot was probed for H218 mRNA (panel A), stripped, and then probed for rat ribosomal RNA (panel B) as an extraction, loading, and transfer control.

FIGS. 5A and 5B shows an X-ray autoradiograph of a Northern blot illustrating the effect of PMA treatment on H218 mRNA levels in RJK88 fibroblasts. Poly-A RNA was extracted from 2 independent 100 mm plates of cells treated with PMA for 2 hrs (PMA) or 2 parallel plates of cells treated with vehicle (CONTROL). The resulting blot was probed for H218 mRNA (panel A), stripped, and then probed for cyclophilin mRNA (panel B) as an extraction, loading, and transfer control. Lanes are presented in pairs based on their relative mRNA content (as indicated by the cyclophilin data).

FIGS. 6A and 6B shows an X-ray autoradiograph of a Northern blot illustrating the effect of NGF treatment on H218 mRNA levels in PC12 cells. Poly-A RNA was extracted from 4 independent 100 mm plates of cells treated with NGF for either 1, 4, or 8 hrs or with a vehicle (CONTROL). The blot was probed for H218 mRNA (panel A), stripped, and then probed for cyclophilin mRNA (panel B) as an extraction, loading, and transfer control.

FIGS. 7A-7C shows the nucleotide (SEQ ID NO.3) and deduced amino acid sequence (SEQ ID NO.4) of rat-edg cDNA. An AITTA motif is boxed in black.

FIG. 8 shows a partial nucleotide sequence (SEQ ID NO.15) of a cDNA that encodes a human p^{H218} polypeptide (SEQ ID NO.16).

BRIEF DESCRIPTION OF THE SEQUENCES

SEQ ID NO. 1 is the nucleotide sequence of the ^{H218} cDNA.

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SEQ ID NO. 2 is the deduced amino acid sequence of the p^{H218} protein encoded by the H218 cDNA.

SEQ ID NO. 3 is the nucleotide sequence of the rat-edg cDNA.

SEQ ID NO. 4 is the deduced amino acid sequence of the p^{rat-edg} protein encoded by the rat-edg cDNA.

SEQ ID NO. 5 is the amino acid sequence of a synthetic p^{H218} peptide designated peptide 1.

SEQ ID NO. 6 is the amino acid sequence of a synthetic p^{H218} peptide designated peptide 2.

SEQ ID NO. 7 is the amino acid sequence of a synthetic p^{H218} peptide designated peptide 3.

SEQ ID NO. 8 is the amino acid sequence of a synthetic p^{H218} peptide designated peptide 4.

SEQ ID NO. 9 is the amino acid sequence of a D2 dopaminergic receptor.

SEQ ID NO. 10 is the amino acid sequence of a β2 adrenergic receptor.

SEQ ID NO. 11 is the amino acid sequence of a α2 adrenergic receptor.

SEQ ID NO. 12 is the amino acid sequence of a 1A serotonergic receptor.

SEQ ID NO. 13 is the amino acid sequence of a M1 muscarinic receptor.

SEQ ID NO. 14 is the amino acid sequence of a substance K receptor.

SEQ ID NO. 15 is a partial nucleotide sequence encoding a human p^{H218} polypeptide.

SEQ ID NO. 16 is an amino acid sequence of a human p^{H218} polypeptide encoded by SEQ ID NO. 15.

DETAILED DISCLOSURE OF THE INVENTION

The subject invention concerns novel cDNAs (H218 and rat-edg) that encode G-protein coupled receptors. The proteins, designated p^{H218} and p^{rat-edg}, play important roles in cell proliferation and differentiation, and in disease states such as cancer.

It has been determined that the protein encoded by H218 polynucleotides is a receptor for sphingosine-1-phosphate (S1P). The research literature indicates that S1P can affect cellular processes potentially involved in many functions including nervous system development, nervous system responses to injury, tumorigenesis, metastasis, inflammation and heart function (Bunemann et al., 1996; Postma et al., 1996; van Koppen et al., 1996; Kawa et al., 1997; Yamamura et al., 1997). Therefore, agonists and antagonists for H218 may be of great clinical value in the treatment of disorders related to the above listed functions, and potentially other, as yet to be discovered, functions.

Rat H218 cDNA has been completely sequenced (SEQ ID NO. 1) and the amino acid sequence of the polypeptide that it encodes determined (SEQ ID NO. 2) (FIG. 1). The H218 cDNA contains a 1056 bp open reading frame that encodes a polypeptide of 352 amino acids. The 3Q untranslated region of H218 cDNA contains repetitive sequences, a consensus sequence for mRNA instability, and a series of terminal adenosines preceded by a potential polyadenylation site. The predicted cytoplasmic regions of p^{H218} contain potential nucleotide binding site components and a consensus sequence for proline directed kinases involved in cell division and growth factor responses.

Analysis of the deduced amino acid sequence of p^{H218} revealed that it is a member of the GPR superfamily (FIG. 2). Several features of p^{H218} are common to all other GPRs,

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including: 1) seven regions of hydrophobicity which are predicted to act as membrane spanning domains, 2) a consensus sequence for N-linked glycosylation in its predicted N-terminal extracellular domain, and 3) a conserved cysteine residue and several serine and threonine residues in its predicted intracellular C-terminal domain. In addition, p^{H218} contains many other residues which are highly conserved among most GPRs. However, p^{H218} is distinct from these GPRs in that it does not contain certain highly conserved residues. Perhaps most notable are the aspartate and tyrosine residues at the cytoplasmic end of the third transmembrane domain, and the cysteine residue at the extracellular end of the same transmembrane domain.

p^{H218} affects the course of cellular proliferation and/or differentiation events. Of all cloned proteins, p^{H218} is most homologous to human p^{edg} , a putative GPR implicated in endothelial cell differentiation. The possibility of a direct interaction between p^{H218} and growth-related intracellular proteins is suggested by the similarity between the predicted cytoplasmic region of p^{H218} and motifs of the src homology domain 2 (SH2) found in many cytoplasmic proteins that are critically involved in growth-related signal transduction, including several proteins encoded by oncogenes.

A further aspect of the subject invention concerns polynucleotide molecules which encode the human homolog of the rat H218 gene. Human cDNAs that hybridize with rat H218 cDNA were isolated from a human embryonic brain cDNA library. A cDNA sequence (SEQ ID NO.15) encoding part of the human H218 protein (SEQ ID NO.16) is shown in FIG. 8. Also contemplated within the scope of the present invention are human genomic H218 polynucleotide sequences, including polynucleotide sequences that flank the protein coding region. These sequences include regulatory sequences and intron sequences.

The human H218 protein is also contemplated within the scope of the invention. In one embodiment, the human H218 protein comprises the amino acid sequence shown in FIG. 8. Fragments and variants of the human H218 protein, including those that are biologically active or that are capable of ligand binding, are also within the scope of the invention.

The subject invention also concerns methods for screening for and identifying ligands of the H218 proteins of the invention. The H218 polynucleotides of the invention can be used to express the H218 protein in any of a wide variety of different expression systems. The H218 protein can then be used to identify H218 agonists and H218 antagonists. The agonists and antagonists can be identified based on their ability to bind to the H218 protein. For example, polynucleotides encoding H218 can be introduced into procaryotic or eucaryotic cells, thereby causing the cells to make H218 protein from the H218 polynucleotides. Therefore, these cells express more H218 protein than they would if the H218 polynucleotides had not been introduced into them. Consequently, H218 ligands can be identified by screening compounds for their ability to bind more to the cells that express greater levels of H218 protein. The ligands may be labeled with radioactive isotopes, or chemical modifications. Alternatively, unmodified ligands may be tracked with other approaches such as antibody recognition. In addition, ligands can also be identified based on their ability to activate H218 protein. When identifying H218 ligands based on their ability to activate H218, activation of H218 can be measured using any of a number of different methods known in the art. For example, one can measure changes in 1) H218-induced intracellular signal transduction events, 2) H218 conformation, 3) proteins bound to H218, and 4) H218-induced changes in cell behavior or morphology.

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Ligands identified using the above methods are also within the scope of the invention.

H218-related nucleic acids can also be used to reduce expression of H218 protein in cells. The ability of ligands to bind to H218 protein or to activate H218, as discussed above, can then be used to identify H218 ligands by comparing cells which express native concentrations of H218 with cells in which H218 concentrations have been reduced. Methods to reduce H218 protein concentrations in cells include, for example, antisense techniques and homologous recombination techniques.

The subject invention also concerns H218-specific nucleic acid probes which can be used to identify mutations in the H218 gene. Identifying such mutations may be important in the scientific study of those diseases which involve mutations in the H218 gene. In addition, identifying the mutations may contribute to the clinical diagnosis, management and counseling related to these diseases. Similarly, antibodies raised against H218 protein sequence may be useful in identifying disease-related changes in H218 protein. The scientific and clinical uses of such antibodies would be the same as those outlined above for the H218-specific nucleic acid probes.

The subject invention also concerns methods for providing gene therapy to a patient in need of such therapy by introducing into the cells of the patient, by in vivo or ex vivo means, a polynucleotide vector that can increase or decrease H218 expression. A polynucleotide expression vector comprising a polynucleotide of the invention that encodes an H218 protein can be used to increase levels of H218 expression in cells. In one embodiment, the polynucleotide encodes the human H218 protein which comprises the amino acid sequence shown in FIG. 8 (seq id no.16). Similarly, H218 protein levels can be decreased in cells using, for example, antisense sequences or homologous recombination techniques. Methods of providing gene therapy are known in the art.

A further aspect of the subject invention concerns antibodies raised against synthetic peptides of p^{H218} . These peptides, designated as 1, 2, 3, and 4 (and corresponding to SEQ ID NO. 5, SEQ ID NO. 6, SEQ ID NO. 7, and SEQ ID NO. 8, respectively), correspond to separate extracellular and intracellular regions of p^{H218} . These peptides and their amino acid sequence are shown in Table 1.

TABLE 1

Amino Acid Sequences of p^{H218} peptides		
p^{H218} peptide	Sequence	
peptide 1	SEQ ID NO. 5	KETLDMQETPSR
peptide 2	SEQ ID NO. 6	YSEYLNPEKVQE
peptide 3	SEQ ID NO. 7	RQGKGATGRRGG
peptide 4	SEQ ID NO. 8	RSSSSLERGLHM

Polyclonal antibodies that react with the antigen peptides were raised in rabbits immunized with the respective peptide. Each antibody recognizes by an ELISA assay the specific peptide used as the immunogen. One of the antibodies, from a rabbit immunized with peptide 1 (SEQ ID NO. 5), was affinity purified and used in a Western blot with antigens from a cell line that expresses H218 mRNA. This antibody recognized a band of 50 to 55 kDa, and a band of 180 to 200 kDa in the Western blot. These antibodies can be used for detecting and purifying the p^{H218} protein through standard procedures known in the art. The antibodies can also be used for localization of p^{H218} in tissues using immunohistochemical techniques known in the art.

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The subject invention further contemplates the use of the protein and peptides to generate both polyclonal and monoclonal antibodies. Contemplated within the scope of the invention are antibodies to wild type forms of H218, as well as mutated forms of H218 protein. Monoclonal antibodies to p^{H218} , and peptide fragments thereof, can be produced using the teachings provided herein in combination with procedures that are well known in the art. Such antibodies can be produced in several host systems, including mouse, rat, and human. In one embodiment, the antibodies of the invention bind to human H218 protein.

Also included within the scope of the invention are binding fragments of the antibodies of the subject invention. Fab', F(ab')₂, and Fv fragments may be obtained by conventional techniques, such as proteolytic digestion of the antibodies by papain or pepsin, or through standard genetic engineering techniques using polynucleotide sequences that encode binding fragments of the antibodies of the subject invention.

A further aspect of the subject invention concerns the cloning and sequencing of the rat homolog of the human edg gene, which also encodes a GPR. This rat gene, designated rat-edg, is similar in sequence to the human edg gene. The rat-edg cDNA (SEQ ID NO. 3) encodes a protein, $p^{rat-edg}$ (SEQ ID NO. 4). The $p^{rat-edg}$ protein also has several features in common with other members of the GPR superfamily including 1) seven hydrophobic regions presumed to act as transmembrane domains, 2) a putative N-glycosylation site in the N-terminal domain, 3) putative phosphorylation sites in cytoplasmic domains, and 4) a conserved cysteine residue in the C-terminal domain.

The subject invention also concerns polynucleotide molecules having sequences that are antisense to mRNA transcripts of H218 and rat-edg polynucleotides. An administration of an antisense polynucleotide molecule can block the production of the protein encoded by H218 or rat-edg. The techniques for preparing antisense polynucleotide molecules, and administering such molecules are known in the art. For example, antisense polynucleotide molecules can be encapsulated into liposomes for fusion with cells.

As is well known in the art, the genetic code is redundant in that certain amino acids are coded for by more than one nucleotide triplet (codon). The subject invention includes those polynucleotide sequences which encode the same amino acids using a different codon from that specifically exemplified in the sequences herein. Such a polynucleotide sequence is referred to herein as an "equivalent" polynucleotide sequence. Thus, the scope of the subject invention includes not only the specific polynucleotide sequences depicted herein, but also all equivalent polynucleotide sequences encoding the polypeptides of the subject invention, and fragments or variants thereof.

The polynucleotide sequences of the subject invention can be prepared according to the teachings contained herein, or by synthesis of oligonucleotide fragments, for example by using a "gene machine" using procedures well known in the art.

The polypeptides of the subject invention can be prepared by expression of the cDNAs in a compatible host cell using an expression vector containing the polynucleotide sequences of the subject invention. The polypeptides can then be purified from the host cell using standard purification techniques that are well known in the art. Alternatively, the polypeptides of the subject invention can be chemically synthesized using solid phase peptide synthesis techniques known in the art.

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The polypeptides of the subject invention can be used as molecular weight markers, as an immunogen for generating antibodies, and as an inert protein in certain assays. The polynucleotide molecules of the subject invention can be used as DNA molecular weight markers, as a chromosome marker, and as a marker for the gene on the chromosome.

The term "polynucleotide sequences" when used in reference to the subject invention can include all or a portion of the cDNA. Similarly, polynucleotide sequences of the subject invention also includes variants, including allelic variations or polymorphisms of the genes. The polynucleotide sequences of the invention may be composed of either RNA or DNA. More preferably, the polynucleotide sequences of the subject invention are composed of DNA.

As used herein, the term "isolated" means, in the case of polynucleotide sequences, that the sequence is no longer linked or associated with other polynucleotide sequences with which it would naturally occur. Thus, the claimed polynucleotide sequences can be inserted into a plasmid or other vector, to form a recombinant DNA cloning vector. The cloning vector may be of bacterial or viral origin. The vector may be designed for the expression of the polypeptide encoded by the polynucleotide sequence. The vector may be transformed or transfected or otherwise inserted into a host cell. The host cell may be either prokaryotic or eukaryotic, and would include bacteria, yeast, insect cells, and mammalian cells. For example, a bacterial host cell may be *E. coli*, and a mammalian host cell may be the PC12 cell line.

As used herein, the term "isolated" means, in the case of proteins, obtaining the protein in a form other than that which occurs in nature. This may be, for example, obtaining p^{H218} by purifying and recovering the protein from a host cell transformed to express the recombinant protein. In the case of antibodies, "isolated" refers to antibodies, which, through the hand of man, have been produced or removed from their natural setting. Thus, isolated antibodies of the subject invention would include antibodies raised as the result of purposeful administration of the proteins, or peptide fragments thereof, of the subject invention in an appropriate host.

The various genetic engineering methods employed herein are well known in the art, and are described in Sambrook, J., et al. (1989) *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory, New York. Thus, it is within the skill of those in the genetic engineering art to screen cDNA libraries, perform restriction enzyme digestions, electrophorese DNA fragments, tail and anneal vector and insert DNA, ligate DNA, transform or transfect host cells, prepare vector DNA, electrophorese proteins, sequence DNA, perform Northern, Southern and Western blotting, and perform PCR techniques.

Materials and Methods

Cloning of H218 cDNA. A "LAMBDA ZAP" cDNA library (Stratagene, La Jolla, Calif.) constructed using rat hippocampal RNA was screened at medium stringency with a 926 bp 5Q EcoRI-Bgl II 3Q fragment of a D2 dopamine receptor cDNA (MacLennan et al., 1990). The cDNA was labeled with ³²P by random hexamer priming. Nitrocellulose filters were incubated for 2 hrs at 42° C. in 5×SSPE (1×SSPE=0.15 M NaCl, 12 mM NaH₂PO₄·H₂O, 1 mM EDTA, pH 7.4), 40% formamide, 0.15% SDS, 5×Denhardt's solution, 100 µg/ml denatured salmon sperm DNA, and 2 µg/ml polyadenylic acid. The filters were then incubated overnight in the same solution at 42° C. with the probe added (approximately 10⁶ cpm/ml). The

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filters were washed two times for 15 minutes each at room temperature in 2×SSC (standard saline citrate buffer: 1×SSC=0.15 M NaCl, 0.015 M sodium citrate, pH 7.2), followed by two washes for 45 minutes each at 42° C. in 2×SSC.

In order to exclude D2 receptor cDNAs from analysis, all hybridizing phage were screened at high stringency with four oligodeoxynucleotide probes designed to specifically recognize D2 dopamine receptor cDNAs (MacLennan et al., 1990). All phage that hybridized to the oligonucleotides were eliminated from further rounds of purification. All other phage that hybridized to the cDNA probe were purified, converted into "BLUESCRIPT" plasmids (Stratagene) according to the manufacturer's automatic excision protocol, and evaluated by restriction digests and gel electrophoresis. Sequence analysis revealed that one of the hybridizing cDNAs, designated "H2", encodes a portion of a putative G-protein coupled receptor (GPR), based on sequence comparisons to other GPRs.

A modified polymerase chain reaction (PCR) technique was used to clone the 5Q cDNA for the H218 cDNA (Loh et al., 1989). H2 cDNA extends 2.6 kb to a 5' end that encodes part of the presumed extracellular N-terminal domain of the receptor. Thus, an oligodeoxynucleotide corresponding to the antisense strand of H2 (nucleotides 288 to 312 of H218) primed the first strand cDNA synthesis with M-MLV Reverse Transcriptase (Gibco-BRL, Gaithersburg, Md.). Poly-A RNA extracted from postnatal day 14 (P14) rat lung served as a template. Terminal Deoxynucleotidyl Transferase (Gibco-BRL) was used to "tail" the resulting cDNA with guanines. The cDNA was then subjected to 35 rounds of PCR amplification with "AMPLITAQ" DNA polymerase (Perkin-Elmer, Branchburg, N.J.). The reaction was primed with an internal H2 specific primer containing antisense strand nucleotides 263 to 288 of H218 and a primer containing a poly-cytosine sequence. The resulting "18" cDNA was subcloned into a "BLUESCRIPT" plasmid (Stratagene) by exploiting restriction sites designed into the 5' ends of the PCR primers.

The "H2" and "18" cDNA fragments were then spliced together to form a 2.75 kb cDNA (designated "H218") containing a complete open reading frame (ORF) of 1052 bp that encodes a polypeptide of 352 amino acids.

Characterization of cDNA Clones The nucleotide sequences of both strands of the H218 cDNA were determined by the dideoxy chain termination technique (Sanger et al., 1977). The T7 Sequencing kit (Pharmacia, Piscataway, N.J.) was used with denatured, double-stranded cDNAs in "BLUESCRIPT" plasmids serving as templates.

Tissue Preparation For RNA preparations, Long Evans rats were killed by decapitation and their brains were immediately removed and dissected. Individual brain regions were frozen in liquid nitrogen. Rats and embryos of both sexes were used in the developmental study. Brains taken from embryos are designated with an "E" and those taken postnatally are designated with a "P". For example, a brain removed 20 days after birth would be P20.

RNA Preparation, Electrophoresis, and Blotting Frozen, dissected brain regions were pooled. The "FASTTRACK" kit (Invitrogen Corp., San Diego, Calif.) was used to extract Poly-A RNA from tissue culture cells and brain tissue used in the developmental study. Total RNA was extracted by homogenization in 4 M guanidine thiocyanate followed by centrifugation through 5.7 M CsCl according to the method of Chirgwin (Chirgwin et al., 1979). The RNA was purified by repeated ethanol precipitations, and its concentration was estimated spec-

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trophotometrically from A₂₆₀. All RNA samples were stored at -20° C. as ethanol precipitates.

RNA (1-10 µg of Poly-A or 20 µg of total) was denatured in 50% deionized formamide, 6.0% formaldehyde at 65° C. for 5 min and then size-fractionated by electrophoresis on a horizontal agarose gel (1.25%) containing 6.0% formaldehyde. The RNA was subsequently transferred to nylon membranes (ICN BIOTRANS membrane), which were then dried and baked at 80° C. for 2 hours under vacuum. Membranes were prehybridized for 2 hrs at 42° C. in 5×SSC, 50% formamide, 0.5% SDS, 50 mM sodium phosphate (pH 6.5) containing 250 µg/ml denatured salmon sperm DNA, 5×Denhardt's solution, and 100 µg/ml polyadenylic acid. The H2 cDNA probe was then ³²P-labeled by random hexamer priming, and added to the prehybridization solution. After hybridization at 42° C. overnight, the membranes were washed twice for 30 min at room temperature in 2×SSC and twice for 45 min at 60° C. in 0.1×SSC, 0.1% SDS.

Membranes were exposed to X-ray film with two intensifying screens at -80° C. for several different time intervals in order to ensure that all comparisons were made within the linear sensitivity range of the film. The probe was then removed from the membranes by washing at 65° C. in 50% formamide, 10 mM sodium phosphate, pH 6.5%, for 1 hour. Stripped blots were rinsed in 2×SSC, 0.1% SDS and exposed to film to check for complete removal of probe. To correct for possible intersample variability in extraction, loading, or transfer of the RNA, the membranes were probed with ³²P-labeled rat cDNA that recognizes ribosomal RNA or with a rat cyclophilin cDNA. Brain Cyclophilin mRNA levels are reported to be stable during brain development (Danielson et al., 1988).

Tissue Culture Cells were grown on plates in Dulbecco's Modified Eagle Media (DMEM) containing 10% fetal bovine serum (FBS), with the exception of PC12 cells which were grown in RPMI media containing 10% horse serum and 5% FBS. Tissue culture cells were washed with 1×PBS, pH 7.4 while anchored to plates, mechanically dislodged, and collected by centrifugation for RNA extraction.

Antibody Production Four peptides having amino acid sequences based on the deduced sequence of p^{H218}, and that correspond to separate extracellular and intracellular regions of p^{H218} were synthesized by the Interdisciplinary Center for Biotechnology Research Core lab at the University of Florida. Rabbits were immunized with the peptides and antiserum prepared according to standard methods. Antisera (designated "1A") from the rabbit immunized with peptide 1 (SEQ ID NO. 5) was purified by precipitation with 4.1 M saturated ammonium sulfate at 25° C. overnight. The precipitate was dissolved in PBS and dialyzed against several changes of PBS. The 0.1A antibody was then affinity purified over a CNBr-Sepharose affinity column (Sigma Chemical, St. Louis, Mo.) to which the peptide 1 (SEQ ID NO. 5) had been attached. Antibody was eluted with 0.1M glycine, pH 2.5.

Western Blotting Crude cellular protein extract or membrane preparations from cell lines that express H218 mRNA were loaded onto a SDS-PAGE gel and electrophoresed. The proteins were then transferred to nitrocellulose paper and reacted with a 1:500 dilution of purified antibody. Rabbit antibody was then detected with a labeled second-step reagent specific for rabbit antibody.

Cloning of the rat-edg cDNA A 1241 bp EcoRI-BamHI fragment of H2 cDNA was labeled with ³²P by random hexamer priming and used to screen approximately 7.5×

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10⁵ cerebellar cDNAs of a rat cerebellar λ -ZAP library at medium stringency. The final hybridization wash was for 45 minutes at 47° C. in 2×SSC. Hybridizing clones were isolated for further evaluation. Purified clones were transferred into “BLUESCRIPT” plasmids (Stratagene) according to the manufacturer’s protocol. Denatured double-stranded plasmids were sequenced by the dideoxy chain termination method (Sanger et al., 1977).

The following are examples which illustrate procedures and processes, including the best mode, for practicing the invention. These examples should not be construed as limiting, and are not intended to be a delineation of all possible modifications to the technique. All percentages are by weight and all solvent mixture proportions are by volume unless otherwise noted.

EXAMPLE 1

Cloning and Sequence Analysis of H218

A rat hippocampal cDNA library was screened at medium stringency with a rat D2 dopamine receptor cDNA. One of the hybridizing cDNAs, designated “H2”, encodes all but a few amino-terminal residues of a novel G-protein coupled receptor. A cDNA, designated “18”, encoding the remaining amino-terminal residues was isolated using a modified PCR technique. The H218 cDNA was prepared from the two independent, overlapping cDNA clones “H2” and “18” which were isolated as described above. The H2 and 18 cDNAs were spliced together to yield a 2.75 kb cDNA containing a complete 1056 bp ORF encoding 352 amino acids. The corresponding gene will be referred to herein as H218, and the encoded GPR protein as p^{H218}. The nucleotide sequence (SEQ ID NO.1) and the amino acid sequence (SEQ ID NO.2) that it encodes are shown in FIG. 1. The series of cytosines at the 5′ end of the clone result from the PCR procedure used to isolate the “18” cDNA. A database search revealed that p^{H218} is clearly a member of the GPR superfamily (FIG. 2).

EXAMPLE 2

H218 mRNA Expression in Brain Tissue

Poly-A RNA was extracted from whole rat brain at multiple stages of development ranging from embryonic day 12 (E12) to postnatal day 80 (P80; adult). A Northern blot of the rat RNA was probed with the complete H2 cDNA. The blot was washed at progressively higher stringencies and exposed to X-ray film after each wash. The autoradiograph revealed an approximately 3.2 kb transcript at all stages of development (FIG. 3). However, H218 mRNA levels are much higher during brain embryogenesis than during later periods of brain development. This pattern indicates that H218 plays a role in cell proliferation and/or differentiation, which is prevalent during brain embryogenesis, rather than in neurotransmission, which is prevalent later in brain development. However, the H218 gene may be involved during all of these processes.

The autoradiographs following the high stringency wash also contain other bands and/or smears, primarily in the E15 and E18 lanes. These signals displayed a preferential reduction in intensity (relative to the 3.2 kb band) during the series of progressively higher stringency washes leading up to the high stringency wash. Therefore, they most likely represent DNA contamination and/or abundant cross hybridizing mRNAs that are related, but not identical, to H218 mRNA. It is also possible that they may partially represent additional ontogenetically regulated H218 transcripts. However, in a smaller scale Northern blot experiment which examined only E15, E18, and P14 brain H218 mRNA, a single 3.2 kb band at E15 and E18 was detected.

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EXAMPLE 3

H218 mRNA Expression in Other Tissue

A Northern blot analysis of total RNA extracted from various organs of the postnatal day 14 (P14) rat was performed. The blot was probed with the H2 cDNA and washed at high stringency. A 3.2 kb H218 mRNA transcript was present in all tissues examined (FIG. 4). The H218 mRNA was most abundant in the lung. Less was found in the kidney, gut, and skin. A very low level of expression was detected in the spleen, brain and liver. This widespread distribution of H218 mRNA expression outside the brain at this stage of development is consistent with p^{H218} role in cell proliferation and/or differentiation.

EXAMPLE 4

H218 mRNA Expression in Cell Lines

Northern blots were performed using poly-A RNA extracted from seven cell lines. The blots were probed with the H2 cDNA, washed at high stringency, and exposed to X-ray film. H218 mRNA was detected in all rodent cell lines examined. Thus, H218 mRNA is synthesized in B104 rat neuroblastoma cells, C6 rat glioma cells, PC12 rat pheochromocytoma cells, NB41A3 mouse neuroblastoma cells, D6P2T rat Schwannoma cells, NIH3T3 mouse fibroblasts, and RJK88 Chinese hamster fibroblasts. In all cases a prominent 3.2 kb band was observed after the high stringency wash, indicating that the sequence and size of the H218 mRNA transcript is highly conserved among mammals. The relative intensity of the band for each cell line is shown in Table 2.

TABLE 2

Relative H218 mRNA concentrations in cell lines	
B104 rat neuroblastoma cells	+++
PC12 rat pheochromocytoma cells	++
C6 rat glioma cells	+++
D6P2T rat Schwannoma cells	++
NB41A3 mouse neuroblastoma cells	+
NIH3T3 mouse fibroblasts	++
RJK88 hamster fibroblasts	++

Of the cells lines and tissue samples examined, H218 mRNA is most abundant in the B104 neuroblastoma cells and the C6 glioma cells. The presence of relatively high concentrations of H218 mRNA in these primitive transformed cells further confirms that the H218 gene is expressed in the early stages of development.

EXAMPLE 5

Manipulation of H218 mRNA Levels Using PMA and Nerve Growth Factor

RJK88 Chinese hamster fibroblasts were grown to approximately 80% confluence in Dulbecco’s Modified Eagle Media (DMEM) containing 10% fetal bovine serum (FBS). The cells were then “serum-deprived” in DMEM containing 0.5% FBS for 2 days and subsequently treated with phorbol 12-myristate 13-acetate (PMA) at a final concentration of 200 ng/ml. Poly-ARNA was extracted 2 hrs after the initiation of PMA treatment. Control RJK88 cells (processed in parallel with PMA treated cells) were grown, serum-deprived, treated with the vehicle for PMA and extracted. A Northern blot performed using the RNA was probed with the H2 cDNA and washed under high stringency conditions. H218 mRNA was undetectable in the serum-deprived, “quiescent” control cells but was clearly present in the cells treated with PMA (FIG. 5).

The nerve growth factor (NGF)-induced differentiation of PC12 rat pheochromocytoma cells from a phenotypic resem-

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bling proliferating, immature adrenal chromaffin cells to a phenotype resembling differentiated sympathetic neurons has been widely employed as a model of neuronal differentiation. A Northern blot was used to determine whether H218 expression in PC12 cells is affected by NGF stimulation. PC12 cells were grown in RPMI media supplemented with 5% FBS and 10% horse serum. The cells were then serum-deprived in RPMI media containing 0.3% FBS and 0.7% horse serum and treated with NGF (50 ng/ml, 2.5 S) 24 hours later. Poly-A RNA was extracted following 1, 4, or 8 hours of the NGF treatment. Control cells (processed in parallel) were treated identically except they received NGF vehicle instead of NGF. A Northern blot using the RNA was probed with the H2 cDNA and washed at high stringency.

NGF treatment rapidly decreases H218 mRNA concentrations in PC12 cells (FIG. 6). H218 mRNA levels (densitometrically quantitated and normalized to cyclophilin mRNA levels) decreased by 39%, 54%, and 33% following NGF treatment of 1, 4, and 8 hours respectively, but returned to normal by 24 hours of continuous NGF treatment. The apparently transient nature of the H218 mRNA decrease in PC12 cells is unlikely the result of any NGF lability given that 1) NGF is a stable compound in solution and 2) PC12 cells treated with NGF that is only replenished every 2 to 3 days (when the media is exchanged) undergo a continuous differentiation which is reversible upon withdrawal of NGF.

EXAMPLE 6

Production and Characterization of Anti-p^{H218} Antibodies

Rabbit antisera against four p^{H218}-derived synthetic peptides and having the amino acid sequences of SEQ ID NOS. 5, 6, 7, and 8, respectively, were prepared. All antisera specifically recognize, with high titers, the appropriate immunogen peptide by ELISA assay. One of the antisera, designated 1A, has been affinity purified. The purified 1A antiserum recognizes two p^{H218} bands on Western blots of cell lines that express H218 mRNA. Both bands were eliminated when the antiserum was preincubated with the antigen peptide but not when it was preincubated with an equal concentration of an irrelevant control peptide.

In addition, the bands were clearly much more intense from a stable cell line that has been engineered to overexpress p^{H218}. The lower (apparent molecular weight of about 50–55 kDa), and weaker, band resulted from monomeric p^{H218} molecules since it roughly corresponds in size to the deduced amino acid sequence encoded by the H218 mRNA open reading frame. The upper (apparent molecular weight of about 180–200 kDa) and more intense band most likely results from an aggregated form of the protein.

The antibody titer in rabbits injected with p^{H218} peptide 1 (SEQ ID NO. 5) rises after the first few injections but drops thereafter, even with continued injections. This unexpected drop was not seen in the rabbits injected with other peptides. It is possible that the drop is the result of the anti-p^{H218} antibodies in the rabbits blocking the function of p^{H218} which, as discussed, may be involved in the cell proliferation events that are required for antibody production.

EXAMPLE 7

Construction and Characterization of Stable Cell Lines with Increased or Decreased Levels of p^{H218}

PC12 cells were transfected with either 1) a vector designed to synthesize H218 mRNA and thereby lead to overexpression of p^{H218}, 2) a vector designed to synthesize antisense H218 mRNA and thereby reduce expression of endogenous PC12 cell p^{H218}, or 3) the empty vector (as a control). Several stable cell lines derived from each condition were isolated and characterized.

Northern blot analyses indicate that all isolated cell lines designed to overexpress H218 mRNA do express additional

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H218 mRNA derived from the transfected DNA. The transfected DNA was designed so that the resulting H218 mRNA would differ in size from mature PC12 cell H218 mRNA and therefore can be easily distinguished. Western blot analysis on one of the lines expressing the most H218 mRNA indicate that this line expressed significantly more p^{H218} than vector transfected control lines.

Nerve growth factor (NGF) and basic fibroblast growth factor (bFGF) cause PC12 cells to differentiate from a phenotype resembling proliferating, immature cells to a phenotype resembling differentiated sympathetic neurons. This system has been extensively studied as a model of neuronal development. The effects of NGF and bFGF on our stable cell lines were examined to determine if manipulating p^{H218} levels affects PC12 cell differentiation. The morphology of the cell lines was qualitatively recorded in two identical experiments by an observer unaware of the identity of the cell lines. The two cell lines overexpressing the most H218 mRNA, including the line shown to overexpress p^{H218}, displayed a significantly less pronounced, growth factor induced change in cell body morphology when compared to vector transfected controls. Cell lines containing only a small amount of additional (exogenous DNA derived) H218 mRNA, including a line which does not detectably overexpress p^{H218} by Western blot analysis, displayed cell morphology changes indistinguishable from vector transfected controls.

Cell lines transfected with the “antisense” vector displayed a significantly more pronounced growth factor induced change in cell body morphology when compared with vector transfected controls. Therefore, increasing p^{H218} levels decreases differentiation while decreasing the expression of p^{H218} increases cell differentiation.

EXAMPLE 8

Cloning of Human H218 Homolog

We have screened a human embryonic brain cDNA library using protocols as described for the cloning of the H218 cDNA and have isolated a cDNA which hybridizes under medium stringency conditions (two 45 minute washes at 42° C. in 2×SSC without formamide) to two non-overlapping fragments of the rat H218 cDNA. The pattern of restriction sites for this novel clone does not match the pattern of restriction sites found with the human edg cDNA clone, and is, therefore, a part of the human homolog of H218.

EXAMPLE 9

Cloning and Sequence Analysis of Rat-edg

A rat cerebellar cDNA library was screened using the H2 cDNA fragment of H218. The largest hybridizing cDNA was completely sequenced (FIG. 7). This 2234 bp cDNA, designated rat-edg (SEQ ID NO.3), contains a 1149 bp ORF preceded by three in-frame stop codons. The cDNA contains an ATTTA motif in its 3Q untranslated region. This motif has been associated with mRNA degradation. The cDNA will subsequently be referred to herein as rat-edg (SEQ ID NO.3) and the encoded protein as p^{rat-edg} (SEQ ID NO.4).

EXAMPLE 10

Expression of Rat-Edg in RNA in Tissue

The same Northern blot described in Example 2 was stripped and reprobed with the rat-edg cDNA. The blot was then washed at high stringency and exposed to X-ray film. Bands corresponding to an approximately 3.2 kb transcript were visible in all brain regions examined on the resulting autoradiograph. This size is close to the reported 3.0 kb size of human-edg. In contrast to H218 mRNA, the 3.2 kb rat-edg mRNA is preferentially expressed in later stages of postnatal development since a continual increase in mRNA expression is observed throughout development, with high-

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est levels detected at P80. The 3.2 kb band observed following the high stringency wash was not the result of the rat-edg cDNA probe cross-hybridizing to H218 mRNA because: 1) the 3.2 kb transcript recognized by rat-edg displays a pattern of expression which is different from that of H218 mRNA, and 2) the in vitro transcribed H218 and rat-edg RNAs are specifically recognized on Northern blots by the appropriate probes.

A second set of generally weaker bands corresponding to a 4.9 kb transcript was also detected using the rat-edg cDNA. The 4.9 kb bands were not preferentially washed off during a series of progressively higher stringency washes and have been observed in multiple independent experiments. Therefore, they probably reflect an alternative rat-edg gene transcript. Interestingly, the expression of the 4.9 kb rat-edg RNA does not display an obvious trend during the developmental stages examined, and at E18, it is more abundant than the 3.2 kb transcript. In addition, the 4.9 kb rat-edg RNA was detected solely in brain RNA samples.

In addition, a Northern blot was performed with total RNA extracted from several regions of adult rat brain. The blot was probed with the rat-edg cDNA, washed at high stringency, and exposed to X-ray film. Rat-edg mRNA was comparably expressed in every region examined (i.e., the frontal cortex, striatum, ventral forebrain, hippocampus, cerebellum, and substantia nigra/ventral tegmental area). The 4.9 kb transcript may be preferentially expressed in the cerebellum, ventral forebrain, and frontal cortex.

The same Northern blot described in Example 3 was stripped and reprobed with the rat-edg cDNA. The blot was washed at high stringency and exposed to X-ray film. At P14, rat-edg mRNA is expressed in the lung (approximately the same concentration as adult brain) and at a much lower concentration in the liver, spleen, and possibly kidney. However, in contrast to H218 mRNA, rat-edg mRNA was not detected in the gut or skin. As noted above, no 4.9 kb bands are detected in any of these regions although they were visible in lanes of the same Northern that were loaded with brain RNA.

EXAMPLE 11

Expression of Rat-Edg RNA in Cell Lines

The Northern blots described in Example 4 were stripped and reprobed with rat-edg cDNA. They were subsequently washed at high stringency and exposed to X-ray film. Like H218 mRNA, rat-edg mRNA is expressed in NIH3T3 cells, C6 rat glioma cells, and rat PC12 pheochromocytoma cells. In contrast to H218 mRNA, rat-edg mRNA was not detected in RJK88 hamster fibroblasts, D6P2T rat Schwannoma cells, NB41A3 mouse neuroblastoma cells, or B104 neuroblastoma cells. Only the 3.2 kb transcript was detected in NIH3T3 and C6 cells, while only the 4.9 kb transcript is detected in PC12 cells.

It should be understood that the examples and embodiments described herein are for illustrative purposes only and that various modifications or changes in light thereof will be

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suggested to persons skilled in the art and are to be included within the scope and purview of this application and the scope of the appended claims.

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

(iii) NUMBER OF SEQUENCES: 16

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

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(A) LENGTH: 2754 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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ATTGGGGCCT CTTGGCTGAT ATCGCTGATT CTGGGTGGCT TGCCCATCCT GGGCTGGAAT	660
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(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 352 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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65          70          75          80
Leu Ala Gly Val Ala Phe Val Ala Asn Thr Leu Leu Ser Gly Pro Val
85          90          95
Thr Leu Ser Leu Thr Pro Leu Gln Trp Phe Ala Arg Glu Gly Ser Ala
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Phe Ile Thr Leu Ser Ala Ser Val Phe Ser Leu Leu Ala Ile Ala Ile
115         120         125
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130         135         140
Ser Cys Arg Met Leu Met Leu Ile Gly Ala Ser Trp Leu Ile Ser Leu
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165         170         175
Glu Ala Cys Ser Thr Val Leu Pro Leu Tyr Ala Lys His Tyr Val Leu
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Cys Val Val Thr Ile Phe Ser Val Ile Leu Leu Ala Ile Val Ala Leu
195         200         205
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Phe Phe Ala Phe Ala Thr Leu Asn Ser Leu Leu Asn Pro Val Ile Tyr
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Thr Trp Arg Ser Arg Asp Leu Arg Arg Glu Val Leu Arg Pro Leu Leu
290 295 300

Cys Trp Arg Gln Gly Lys Gly Ala Thr Gly Arg Arg Gly Gly Asn Pro
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Gly His Arg Leu Leu Pro Leu Arg Ser Ser Ser Ser Leu Glu Arg Gly
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Leu His Met Pro Thr Ser Pro Thr Phe Leu Glu Gly Asn Thr Val Val
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(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2232 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(ix) FEATURE:

- (A) NAME/KEY: CDS
 (B) LOCATION: 269..1420

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

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AAG CAC TAT ATT CTC TTC TGC ACC ACC GTC TTC ACC CTG CTC CTG CTT	916
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GGAAAACATA GTGCTGAATG ACGGCAAAGA ATGGTGGTAA ATCAAAAGAT AAATTAACCTT 1867
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 AATCAGACAT TTCAGATGCC CGTCAATGTA AAATCACCTA CTTGAACATT GTATGCAATA 1987
 CATTACACA AAAAAGCAAA TACTGTAGCC TTATTTGAAC AATACTGAAC TCATAAATAC 2047
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(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 383 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Val Ser Ser Thr Ser Ile Pro Val Val Lys Ala Leu Arg Ser Gln
 1 5 10 15
 Val Ser Asp Tyr Gly Asn Tyr Asp Ile Ile Val Arg His Tyr Asn Tyr
 20 25 30
 Thr Gly Lys Leu Asn Ile Gly Val Glu Lys Asp His Gly Ile Lys Leu
 35 40 45
 Thr Ser Val Val Phe Ile Leu Ile Cys Cys Leu Ile Ile Leu Glu Asn
 50 55 60
 Ile Phe Val Leu Leu Thr Ile Trp Lys Thr Lys Lys Phe His Arg Pro
 65 70 75 80
 Met Tyr Tyr Phe Ile Gly Asn Leu Ala Leu Ser Asp Leu Leu Ala Gly
 85 90 95
 Val Ala Tyr Thr Ala Asn Leu Leu Ser Gly Ala Thr Thr Tyr Lys
 100 105 110
 Leu Thr Pro Ala Gln Trp Phe Leu Arg Glu Gly Ser Met Phe Val Ala
 115 120 125
 Leu Ser Ala Ser Val Phe Ser Leu Leu Ala Ile Ala Ile Glu Arg Tyr
 130 135 140
 Ile Thr Met Leu Lys Met Lys Leu His Asn Gly Ser Asn Ser Ser Arg
 145 150 155 160
 Ser Phe Leu Leu Ile Ser Ala Cys Trp Val Ile Ser Leu Ile Leu Gly
 165 170 175
 Gly Leu Pro Ile Met Gly Trp Asn Cys Ile Ser Ser Leu Ser Ser Cys
 180 185 190
 Ser Thr Val Leu Pro Leu Tyr His Lys His Tyr Ile Leu Phe Cys Thr
 195 200 205
 Thr Val Phe Thr Leu Leu Leu Ser Ile Val Ile Leu Tyr Cys Arg
 210 215 220
 Ile Tyr Ser Leu Val Arg Thr Arg Ser Arg Arg Leu Thr Phe Arg Lys
 225 230 235 240
 Asn Ile Ser Lys Ala Ser Arg Ser Ser Glu Lys Ser Leu Ala Leu Leu
 245 250 255
 Lys Thr Val Ile Ile Val Leu Ser Val Phe Ile Ala Cys Trp Ala Pro
 260 265 270

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Leu	Phe	Ile	Leu	Leu	Leu	Leu	Asp	Val	Gly	Cys	Lys	Ala	Lys	Thr	Cys
		275					280					285			
Asp	Ile	Leu	Tyr	Lys	Ala	Glu	Tyr	Phe	Leu	Val	Leu	Ala	Val	Leu	Asn
	290					295					300				
Ser	Gly	Thr	Asn	Pro	Ile	Ile	Tyr	Thr	Leu	Thr	Asn	Lys	Glu	Met	Arg
305					310					315					320
Arg	Ala	Phe	Ile	Arg	Ile	Ile	Ser	Cys	Cys	Lys	Cys	Pro	Asn	Gly	Asp
				325					330					335	
Ser	Ala	Gly	Lys	Phe	Lys	Arg	Pro	Ile	Ile	Pro	Gly	Met	Glu	Phe	Ser
			340					345					350		
Arg	Ser	Lys	Ser	Asp	Asn	Ser	Ser	His	Pro	Gln	Lys	Asp	Asp	Gly	Asp
		355					360					365			
Asn	Pro	Glu	Thr	Ile	Met	Ser	Ser	Gly	Asn	Val	Asn	Ser	Ser	Ser	
370						375					380				

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 12 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Lys Glu Thr Leu Asp Met Gln Glu Thr Pro Ser Arg
1 5 10

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 12 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Tyr Ser Glu Tyr Leu Asn Pro Glu Lys Val Gln Glu
1 5 10

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 12 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Arg Gln Gly Lys Gly Ala Thr Gly Arg Arg Gly Gly
1 5 10

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 12 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

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Arg Ser Ser Ser Ser Leu Glu Arg Gly Leu His Met
1 5 10

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 303 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: Not Relevant
- (D) TOPOLOGY: Not Relevant

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Met Asp Pro Leu Asn Leu Ser Trp Tyr Asp Asp Asp Leu Glu Arg Gln
1 5 10 15

Asn Trp Ser Arg Pro Phe Asn Gly Ser Glu Gly Lys Ala Asp Arg Pro
20 25 30

His Tyr Asn Tyr Tyr Ala Met Leu Leu Thr Leu Leu Ile Phe Ile Ile
35 40 45

Val Phe Gly Asn Val Leu Val Cys Met Ala Val Ser Arg Glu Lys Ala
50 55 60

Leu Gln Thr Thr Thr Asn Tyr Leu Ile Val Ser Leu Ala Val Ala Asp
65 70 75 80

Leu Leu Val Ala Thr Leu Val Met Pro Trp Val Val Tyr Leu Glu Val
85 90 95

Val Gly Glu Trp Lys Phe Ser Arg Ile His Cys Asp Ile Phe Val Thr
100 105 110

Leu Asp Val Met Met Cys Thr Ala Ser Ile Leu Asn Leu Cys Ala Ile
115 120 125

Ser Ile Asp Arg Tyr Thr Ala Val Ala Met Pro Met Leu Tyr Asn Thr
130 135 140

Arg Tyr Ser Ser Lys Arg Arg Val Thr Val Met Ile Ala Ile Val Trp
145 150 155 160

Val Leu Ser Phe Thr Ile Ser Cys Pro Leu Leu Phe Gly Leu Asn Asn
165 170 175

Thr Asp Gln Asn Glu Cys Ile Ile Ala Asn Pro Ala Phe Val Val Tyr
180 185 190

Ser Ser Ile Val Ser Phe Tyr Val Pro Phe Ile Val Thr Leu Leu Val
195 200 205

Tyr Ile Lys Ile Tyr Ile Val Leu Arg Lys Arg Arg Lys Arg Val Asn
210 215 220

Thr Lys Lys Glu Lys Lys Ala Thr Gln Met Leu Ala Ile Val Leu Gly
225 230 235 240

Val Phe Ile Ile Cys Trp Leu Pro Phe Phe Ile Thr His Ile Leu Asn
245 250 255

Ile His Cys Asp Cys Asn Ile Pro Pro Val Leu Tyr Ser Ala Phe Thr
260 265 270

Trp Leu Gly Tyr Val Asn Ser Ala Val Asn Pro Ile Ile Tyr Thr Thr
275 280 285

Phe Asn Ile Glu Phe Arg Lys Ala Phe Met Lys Ile Leu His Cys
290 295 300

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 377 amino acids
- (B) TYPE: amino acid

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[illegible]

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(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 450 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: Not Relevant

(D) TOPOLOGY: Not Relevant

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Met 1	Gly	Ser	Leu	Gln 5	Pro	Asp	Ala	Gly	Asn 10	Ala	Ser	Trp	Asn	Gly 15	Thr
Glu	Ala	Pro	Gly 20	Gly	Gly	Ala	Arg	Ala 25	Thr	Pro	Tyr	Ser	Leu 30	Gln	Val
Thr	Leu	Thr 35	Leu	Val	Cys	Leu	Ala 40	Gly	Leu	Leu	Met 45	Leu	Thr	Val	
Phe 50	Gly	Asn	Val	Leu	Val	Ile 55	Ile	Ala	Val	Phe 60	Thr	Ser	Arg	Ala	Leu
Lys 65	Ala	Pro	Gln	Asn	Leu 70	Phe	Leu	Val	Ser 75	Leu	Ala	Ser	Ala	Asp	Ile 80
Leu	Val	Ala	Thr 85	Leu	Val	Ile	Pro	Phe	Ser 90	Leu	Ala	Asn	Glu 95	Val	Met
Gly	Tyr	Trp 100	Tyr	Phe	Gly	Lys	Thr	Trp 105	Cys	Glu	Ile	Tyr 110	Leu	Ala	Leu
Asp 115	Val	Leu	Phe	Cys	Thr	Ser	Ser 120	Ile	Val	His	Leu 125	Cys	Ala	Ile	Ser
Leu 130	Asp	Arg	Tyr	Trp	Ser 135	Ile	Thr	Gln	Ala	Ile 140	Glu	Tyr	Asn	Leu	Lys
Arg 145	Thr	Pro	Arg	Arg 150	Ile	Lys	Ala	Ile	Ile 155	Thr	Val	Trp	Val	Ile 160	
Ser	Ala	Val	Ile 165	Ser	Phe	Pro	Pro	Leu	Ile 170	Ser	Ile	Glu	Lys	Lys 175	Gly
Gly	Gly	Gly 180	Gly	Pro	Gln	Pro	Ala	Glu 185	Pro	Arg	Cys	Glu 190	Ile	Asn	Asp
Gln 195	Lys	Trp	Tyr	Val	Ile	Ser	Ser 200	Cys	Ile	Gly	Ser 205	Phe	Phe	Ala	Pro
Cys 210	Leu	Ile	Met	Ile	Leu	Val 215	Tyr	Val	Arg	Ile 220	Tyr	Gln	Ile	Ala	Lys
Arg 225	Arg	Thr	Arg	Val	Xaa 230	Xaa	Xaa	Xaa	Xaa 235	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa
Xaa	Xaa	Xaa	Xaa 245	Xaa	Xaa	Xaa	Xaa	Xaa 250	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa
Xaa	Xaa	Xaa 260	Xaa	Xaa	Xaa	Xaa	Xaa 265	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa
Xaa	Xaa	Xaa 275	Xaa	Xaa	Xaa	Xaa	Xaa 280	Xaa	Xaa	Xaa	Xaa	Xaa 285	Xaa	Xaa	Xaa
Xaa 290	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa 295	Xaa	Xaa	Xaa	Xaa 300	Xaa	Xaa	Xaa	Xaa
Xaa 305	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa 310	Xaa	Xaa	Xaa	Xaa 315	Xaa	Xaa	Xaa	Xaa
Xaa	Xaa	Xaa	Xaa 325	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa 330	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa
Xaa	Xaa	Xaa 340	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa 345	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa

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(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 421 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: Not Relevant
- (D) TOPOLOGY: Not Relevant

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Met	Asp	Val	Leu	Ser	Pro	Gly	Gly	Asn	Asn	Thr	Thr	Ser	Pro	Pro	Ala
1				5					10					15	
Pro	Phe	Glu	Thr	Gly	Gly	Asn	Thr	Thr	Gly	Ile	Ser	Asp	Val	Thr	Val
			20					25					30		
Ser	Tyr	Gln	Val	Ile	Thr	Ser	Leu	Leu	Gly	Thr	Leu	Ile	Phe	Cys	
		35					40				45				
Ala	Val	Leu	Gly	Asn	Ala	Cys	Val	Val	Ala	Ala	Ile	Ala	Leu	Glu	Arg
		50				55					60				
Ser	Leu	Gln	Asn	Val	Ala	Asn	Tyr	Leu	Ile	Gly	Ser	Leu	Ala	Val	Thr
65					70					75					80
Asp	Leu	Met	Val	Ser	Val	Leu	Val	Leu	Pro	Met	Ala	Ala	Leu	Tyr	Gln
				85					90					95	
Val	Leu	Asn	Lys	Trp	Thr	Leu	Gly	Gln	Val	Thr	Cys	Asp	Leu	Phe	Ile
			100					105						110	
Ala	Leu	Asp	Val	Leu	Cys	Cys	Thr	Ser	Ser	Ile	Leu	His	Leu	Cys	Ala
		115					120					125			
Ile	Ala	Leu	Asp	Arg	Tyr	Trp	Ala	Ile	Thr	Asp	Pro	Ile	Asp	Tyr	Val
		130				135					140				
Asn	Lys	Arg	Thr	Pro	Arg	Pro	Arg	Ala	Leu	Thr	Ser	Leu	Thr	Trp	Leu
145					150					155					160
Ile	Gly	Phe	Leu	Ile	Ser	Ile	Pro	Pro	Met	Leu	Gly	Trp	Arg	Thr	Pro
				165					170					175	
Glu	Asp	Arg	Ser	Asp	Pro	Asp	Ala	Cys	Thr	Ile	Ser	Lys	Asp	Met	Gly
			180					185					190		
Tyr	Thr	Ile	Tyr	Ser	Thr	Phe	Gly	Ala	Phe	Tyr	Ile	Pro	Leu	Leu	Leu
		195					200					205			
Met	Leu	Val	Leu	Tyr	Gly	Arg	Ile	Phe	Arg	Ala	Ala	Arg	Phe	Arg	Ile
		210				215					220				
Pro	Lys	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa
225					230					235					240

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Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
 245 250 255
 Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
 260 265 270
 Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
 275 280 285
 Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
 290 295 300
 Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
 305 310 315 320
 Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
 325 330 335
 Xaa Arg Glu Arg Lys Thr Val Lys Thr Leu Gly Ile Ile Met Gly Thr
 340 345 350
 Phe Ile Leu Cys Trp Leu Pro Phe Phe Ile Val Ala Leu Val Leu Pro
 355 360 365
 Phe Cys Glu Ser Ser Cys His Met Pro Thr Leu Leu Gly Ala Ile Ile
 370 375 380
 Asn Trp Leu Gly Tyr Ser Asn Ser Leu Leu Asn Pro Val Ile Tyr Ala
 385 390 395 400
 Tyr Phe Asn Lys Asp Phe Gln Asn Ala Phe Lys Lys Ile Ile Lys Cys
 405 410 415
 Xaa Xaa Xaa Xaa Xaa
 420

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 461 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: Not Relevant
 (D) TOPOLOGY: Not Relevant

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Met Asn Thr Ser Ala Pro Pro Ala Val Ser Pro Asn Ile Thr Val Leu
 1 5 10 15
 Ala Pro Gly Lys Gly Pro Trp Gln Val Ala Phe Ile Gly Ile Thr Thr
 20 25 30
 Gly Leu Leu Ser Leu Ala Thr Val Thr Gly Asn Leu Leu Val Ile Ile
 35 40 45
 Ser Phe Lys Val Asn Thr Glu Leu Lys Thr Val Asn Asn Tyr Phe Leu
 50 55 60
 Leu Ser Leu Ala Cys Ala Asp Leu Ile Ile Gly Thr Phe Ser Met Asn
 65 70 75 80
 Leu Tyr Thr Thr Tyr Leu Leu Met Gly His Trp Ala Leu Gly Thr Leu
 85 90 95
 Ala Cys Asp Leu Trp Leu Ala Leu Asp Tyr Val Ala Ser Asn Ala Ser
 100 105 110
 Val Met Asn Leu Leu Leu Ile Ser Phe Asp Arg Tyr Phe Ser Val Thr
 115 120 125
 Arg Pro Leu Ser Tyr Arg Ala Lys Arg Thr Pro Arg Arg Ala Ala Leu
 130 135 140
 Met Ile Gly Leu Ala Trp Leu Val Ser Phe Val Leu Trp Ala Pro Ala
 145 150 155 160

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Ile Leu Phe Trp Gln Tyr Leu Val Gly Glu Arg Thr Val Leu Ala Gly
      165      170      175
Gln Cys Tyr Ile Gln Phe Leu Ser Gln Pro Ile Ile Thr Phe Gly Thr
      180      185      190
Ala Met Ala Ala Phe Tyr Leu Pro Val Thr Val Met Cys Thr Leu Tyr
      195      200      205
Trp Arg Ile Tyr Arg Glu Thr Glu Asn Arg Ala Arg Glu Xaa Xaa Xaa
      210      215      220
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
      225      230      235      240
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
      245      250      255
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
      260      265      270
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
      275      280      285
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
      290      295      300
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
      305      310      315      320
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
      325      330      335
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
      340      345      350
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Lys Glu Lys Lys Ala Ala Arg Thr Leu
      355      360      365
Ser Ala Ile Leu Leu Ala Phe Ile Val Thr Trp Thr Pro Tyr Asn Ile
      370      375      380
Met Val Leu Val Ser Thr Phe Cys Lys Asp Cys Val Pro Glu Thr Leu
      385      390      395      400
Trp Glu Leu Gly Tyr Trp Leu Cys Tyr Val Asn Ser Thr Ile Asn Pro
      405      410      415
Met Cys Tyr Ala Leu Cys Asn Lys Ala Phe Arg Asp Thr Phe Arg Leu
      420      425      430
Leu Leu Leu Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
      435      440      445
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
      450      455      460

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(2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 387 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: Not Relevant
 - (D) TOPOLOGY: Not Relevant

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

```

Met Gly Ala Cys Val Val Met Thr Asp Ile Asn Ile Ser Ser Gly Leu
 1      5      10      15
Asp Ser Asn Ala Thr Gly Ile Thr Ala Phe Ser Met Pro Gly Trp Gln
      20      25      30
Leu Ala Leu Trp Thr Ala Ala Tyr Leu Ala Leu Val Leu Val Ala Val
      35      40      45

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Met	Gly	Asn	Ala	Thr	Val	Ile	Trp	Ile	Ile	Leu	Ala	His	Gln	Arg	Met
50						55				60					
Arg	Thr	Val	Thr	Asn	Tyr	Phe	Ile	Val	Asn	Leu	Ala	Leu	Ala	Asp	Leu
65				70					75					80	
Cys	Met	Ala	Ala	Phe	Asn	Ala	Ala	Phe	Asn	Phe	Val	Tyr	Ala	Ser	His
				85					90					95	
Asn	Ile	Trp	Tyr	Phe	Gly	Arg	Ala	Phe	Cys	Tyr	Phe	Gln	Asn	Leu	Phe
			100					105					110		
Pro	Ile	Thr	Ala	Met	Phe	Val	Ser	Ile	Tyr	Ser	Met	Thr	Ala	Ile	Ala
			115				120					125			
Ala	Asp	Arg	Tyr	Met	Ala	Ile	Val	His	Pro	Phe	Gln	Pro	Arg	Leu	Ser
			130			135					140				
Ala	Pro	Gly	Thr	Arg	Ala	Val	Ile	Ala	Gly	Ile	Trp	Leu	Val	Ala	Leu
145				150					155					160	
Ala	Leu	Ala	Phe	Pro	Gln	Cys	Phe	Tyr	Ser	Thr	Ile	Thr	Thr	Asp	Glu
			165					170						175	
Gly	Ala	Thr	Lys	Cys	Val	Val	Ala	Trp	Pro	Glu	Asp	Ser	Gly	Gly	Lys
			180					185					190		
Met	Leu	Leu	Leu	Tyr	His	Leu	Ile	Val	Ile	Ala	Leu	Ile	Tyr	Phe	Leu
		195				200						205			
Pro	Leu	Val	Val	Met	Phe	Val	Ala	Tyr	Ser	Val	Ile	Gly	Leu	Thr	Leu
		210				215					220				
Trp	Arg	Arg	Ser	Val	Pro	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa
225				230					235					240	
Xaa	Xaa	Xaa	Ala	Lys	Lys	Lys	Phe	Val	Lys	Thr	Met	Val	Leu	Val	Val
			245						250				255		
Val	Thr	Phe	Ala	Ile	Cys	Trp	Leu	Pro	Tyr	His	Leu	Tyr	Phe	Ile	Leu
		260				265						270			
Gly	Thr	Phe	Gln	Glu	Asp	Ile	Tyr	Cys	His	Lys	Phe	Ile	Gln	Gln	Val
		275				280					285				
Tyr	Leu	Ala	Leu	Phe	Trp	Leu	Ala	Met	Ser	Ser	Thr	Met	Tyr	Asn	Pro
		290				295					300				
Ile	Ile	Tyr	Cys	Cys	Leu	Asn	His	Arg	Phe	Arg	Ser	Gly	Phe	Arg	Leu
305					310					315				320	
Ala	Phe	Arg	Cys	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa
			325						330					335	
Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa
			340					345					350		
Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa
		355				360					365				
Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa	Xaa
		370				375				380					
Xaa	Xaa	Xaa													
385															

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 162 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

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CCGCAGACGC TAGCCCTGCT CAAGACGGTC ACCATCGTGC TAGGCGTCTT TATCGTCTGC	60
TGGCTGCCCG CCTTCAGCAT CTCCTTCTG GACTATGCCT GTCCCGTCCA CTCCTGCCCG	120
ATCCTCTACA AAGCCCACTA CTTTTTCGCC GTCTCCACCC TG	162

(2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 54 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:
Pro Gln Thr Leu Ala Leu Leu Lys Thr Val Thr Ile Val Leu Gly Val
5 10 15
Phe Ile Val Cys Trp Leu Pro Ala Phe Ser Ile Leu Leu Leu Asp Tyr
20 25 30
Ala Cys Pro Val His Ser Cys Pro Ile Leu Tyr Lys Ala His Tyr Phe
35 40 45
Phe Ala Val Ser Thr Leu
50

I claim:

1. An isolated polynucleotide molecule selected from the group consisting of a polynucleotide which encodes a polypeptide comprising the amino acid sequence shown in SEQ ID NO. 16, and a polynucleotide which is complementary to a polynucleotide which encodes a polypeptide comprising the amino acid sequence shown in SEQ ID NO. 16.

2. An isolated polynucleotide comprising SEQ ID NO. 15.

* * * * *